



Ministry of Community Development, Mother
and Child Health

PMI | Africa IRS (AIRS) Project

Indoor Residual Spraying (IRS Task Order 6)

ZAMBIA 2015 ENTOMOLOGY REPORT

AUGUST 2015- FEBRUARY 2016

Recommended Citation: The President's Malaria Initiative (PMI)/Africa Indoor Residual Spraying Project. August 2016. Zambia Entomology Report, 2015. Bethesda, MD. PMI Africa IRS (AIRS) Project Indoor Residual Spraying (IRS 2) Task Order Six, Abt Associates Inc.

Contract No. and Task order: GHN-I-00-09-00013 & AID- OAA-TO-14-00035

Submitted to: United States Agency for International Development/PMI

Prepared by: Abt Associates Inc.

Submitted on: June 17, 2016

Re-Submitted on: August 5, 2016

Re-Submitted on: August 25, 2016

Re-Submitted and Approved: September 8, 2016



Abt Associates Inc. 1 4550 Montgomery Avenue 1 Suite 800 North

1 Bethesda, Maryland 20814 1 T. 301.347.5000 1 F. 301.913.9061

ACRONYMS

AIRS	Africa Indoor Residual Spraying
CDC	Centers for Disease Control and Prevention
CSP	Circumsporozoite Antigens
DDMS	Disease Data Management System
DDT	Dichlorodiphenyltrichloroethane
EIR	Entomological Inoculation Rate
ELISA	Enzyme-linked Immunosorbent Assay
HLC	Human Landing Catch
I-RDT	Intensity Rapid Diagnostic Test
IRS	Indoor Residual Spraying
PSC	Pyrethrum Spray Catch
PCR	Polymerase Chain Reaction
PMI	President's Malaria Initiative
WHO	World Health Organization
WHOPES	World Health Organization Pesticides Evaluation Scheme

CONTENTS

Acronyms	3
Executive Summary	8
1. Introduction	10
2. Materials and Methods	12
2.1 Study sites	12
2.2 Malaria Vector Density and Behavior Monitoring	13
2.2.1 Pyrethrum Spray Catch (PSC).....	13
2.2.2 CDC light trap collections	13
2.2.3 Human Landing Catch (HLC).....	14
2.3 Susceptibility tests	14
2.3.1 WHO susceptibility test.....	14
2.3.2 CDC Resistance Intensity Rapid Diagnostic Test (I-RDT)	15
2.4 Quality assurance of IRS and insecticide decay rate.....	15
3. Results and Discussion	17
3.1 Species composition, vector density, behavior and longevity.....	17
3.1.1 Species composition	17
3.2 Vector density, behavior and longevity	18
3.2.1 Pyrethrum Spray Catch	18
3.2.2 CDC light trap collection.....	24
3.2.3 HLC Collection	29
3.3 <i>Anopheles gambiae</i> s.l. biting rate.....	37
3.4 <i>Anopheles gambiae</i> s.l. biting time.....	41
3.5 Parity rate.....	43
3.6 Quality Assurance of IRS and Insecticide Decay Rate	45
3.7 Malaria vector susceptibility tests	46
4. Training on the Disease Data Management System (DDMS)	54
5. County Capacity Building	55

Annex	58
-------------	----

LIST OF FIGURES

Figure 1: Geographical locations of entomological sentinel sites in Zambia	13
Figure 2: Species composition in the targeted sprayed sentinel sites (Aug 2015 - Jan 2016)	17
Figure 3 : Species composition in the control sentinel sites (Aug 2015 - Jan 2016).....	18
Figure 4: Indoor resting density of <i>Anopheles funestus</i> s.l. per room per night in August (baseline) and one, two and three months after IRS.....	21
Figure 5: Average density of <i>Anopheles gambiae</i> s.l. per room per day in Mwense District (Aug 2015 - Jan 2016).....	21
Figure 6: Average density of <i>Anopheles gambiae</i> s.l. per room per day in Milenge District (Aug 2015 - Jan 2016).....	22
Figure 7: Average density of <i>Anopheles gambiae</i> s.l. per room per day in Kasama and Isoka Districts (Aug 2015 - Jan 2016)	22
Figure 8: Average density of <i>Anopheles funestus</i> s.l. per trap per night in Katete District (Aug 2015 - Jan 2016).....	24
Figure 9: Average density of <i>Anopheles funestus</i> s.l. per trap per night in Mwense District (Aug 2015 - Jan 2016).....	25
Figure 10: Average density of <i>Anopheles gambiae</i> s.l. per trap per night in Mwense District (Aug 2015 - Jan 2016).....	25
Figure 11 : Average density of <i>Anopheles funestus</i> s.l. per trap per night in Milenge District (Aug 2015 - Jan 2016)	26
Figure 12: Average density of <i>Anopheles gambiae</i> s.l. per trap per night in Milenge District (Aug 2015 - Jan 2016).....	27
Figure 13: Average density of <i>Anopheles funestus</i> s.l. per trap per night in Serenje District (Aug 2015 - Jan 2016).....	27
Figure 14: Average density of <i>Anopheles funestus</i> s.l. per trap per night in Kasama and Isoka districts (Aug 2015 - Jan 2016).....	28
Figure 15: Average density of <i>Anopheles gambiae</i> s.l. per trap per night in Katete District from August 2015 to January 2016.....	29
Figure 16: <i>Anopheles</i> species composition of the indoor HLC (Aug 2015 to Jan 2016).....	30
Figure 17: <i>Anopheles</i> species composition of the outdoor HLC (Aug 2015 to Jan 2016) ...	30
Figure 18: <i>Anopheles funestus</i> s.l. feeding time in Lunga in Milenge District, before and after IRS.....	35
Figure 19: <i>Anopheles funestus</i> s.l. feeding time in Shibesa in Mwense District after IRS ...	36

Figure 20: <i>Anopheles funestus</i> s.l. feeding time in Chibobo in Serenje district before and after IRS.....	36
Figure 21: <i>Anopheles funestus</i> s.l. feeding time in Kalonga in Kasama District before and after IRS.....	36
Figure 22: <i>Anopheles funestus</i> s.l. feeding time in Nsalamba in Isoka District before and after IRS.....	37
Figure 23: <i>Anopheles gambiae</i> s.l. feeding time in Lunga (sprayed site) in Milenge	41
Figure 24: <i>Anopheles gambiae</i> s.l. feeding time in Shibesa (sprayed site) in Mwense	41
Figure 25: <i>Anopheles gambiae</i> s.l. feeding time in Kalonga (sprayed site) in Kasama	42
Figure 26: <i>Anopheles gambiae</i> s.l. feeding time in Nsalamba (sprayed site) in Isoka.....	42
Figure 27: Mortality of Kisumu susceptible strain of <i>An. gambiae</i> s.s. after 30 mins exposure to pirimiphos-methyl CS and 24H holding period at T0, T1, T2, T3, and T4 in Kasama, Isoka, and Katete	46
Figure 28: Mortality of <i>Anopheles funestus</i> s.l. susceptible strain after 30 mins exposure to pirimiphos-methyl CS and 24H holding period at T0, T1, T2, T3 and T4	46
Figure 29: Susceptibility status of <i>Anopheles funestus</i> s.l.	47
Figure 30 : Susceptibility status of <i>Anopheles gambiae</i> s.l.....	47
Figure 31: Intensity resistance assay in Milenge district against <i>An. funestus</i> s.l.	48
Figure 32: Intensity resistance assay in Samfya District	52

LIST OF TABLES

Table 1: Entomological surveillance sentinel sites.....	12
Table 2 : Blood digestion stage and average density of <i>An. funestus</i> s.l. collected by PSC per room in Serenje District in August and from November 2015 to January 2016	19
Table 3 : Blood digestion stage and average density of <i>An. funestus</i> s.l. collected by PSC per room in Milenge District in August and from November 2015 to January 2016.....	19
Table 4 : Blood digestion stage and average density of <i>An. funestus</i> s.l. collected by PSC per room in Mwense District in August and from November 2015 to January 2016.....	20
Table 5 : Blood digestion stage and average density of <i>An. funestus</i> s.l. collected by PSC per room in Kasama and Isoka districts in August and from November 2015 to January 2016	20
Table 6: <i>Anopheles funestus</i> s.l. HBR in sprayed and control sites in August and from November 2015 to January 2016	31
Table 7: <i>Anopheles funestus</i> s.l. HBR in Katete District in August and from November 2015 to January 2016	32
Table 8: <i>Anopheles funestus</i> s.l. HBR in Mwense District in August and from November 2015 to January 2016	32

Table 9 : <i>Anopheles funestus</i> s.l. HBR in Milenge District in August and from November 2015 to January 2016	33
Table 10: <i>Anopheles funestus</i> s.l. HBR in Mwense District in August and from November 2015 to January 2016	34
Table 11: <i>Anopheles funestus</i> s.l. biting rate in Kasama and Isoka districts in August and from November 2015 to January 2016.....	34
Table 12: <i>Anopheles gambiae</i> s.l. biting rate in Milenge District in August and from November 2015 to January 2016	38
Table 13: <i>Anopheles gambiae</i> s.l. biting rate in Mwense District in August and from November 2015 to January 2016	39
Table 14: <i>Anopheles gambiae</i> s.l. biting rate in Serenje District in August and from November 2015 to January 2016	39
Table 15: <i>Anopheles gambiae</i> s.l. biting rate in Kasama and Isoka districts in August and from November 2015 to January 2016.....	40
Table 16: <i>Anopheles funestus</i> s.l. parity rate in Milenge and Mwense districts	43
Table 17: <i>Anopheles funestus</i> s.l. parity rate in Serenje and Katete districts	43
Table 18: <i>Anopheles funestus</i> s.l. parity rate in Kasama and Isoka districts	43
Table 19: <i>Anopheles gambiae</i> s.l. parity rate in Milenge and Mwense districts	44
Table 20: <i>Anopheles gambiae</i> s.l. parity rate in Serenje and Katete districts.....	44
Table 21: <i>Anopheles gambiae</i> s.l. parity rate in Kasama and Isoka districts	44

EXECUTIVE SUMMARY

Zambia is implementing indoor residual spraying (IRS) for malaria control as part of an integrated vector management strategy. The President's Malaria Initiative (PMI) Africa Indoor Residual Spraying (AIRS) Project in Zambia conducted indoor residual spraying (IRS) in 39 districts in 2015 from September 28 – November 25, 2015. Entomological surveillance was carried out in six sprayed targeted sites and four control sites.

Methods:

Baseline data were collected in August 2015 using pyrethrum spray catch (PSC), Centers for Disease Control and Prevention (CDC) light trap and human landing catch (HLC). The impact of the IRS campaign on the malaria vectors was assessed from November 2015 to February 2016. The quality assurance of the IRS operations was assessed 24h after the spraying and the assessment of decay rate of insecticide sprayed was followed on a monthly basis. The level of the resistance of *Anopheles funestus* s.l. and *Anopheles gambiae* s.l. was assessed using the World Health Organization (WHO) tube tests. The resistance intensity assays was also performed in Mwenze, Milenge, and Samfya districts of Luapula Province using the CDC bottle assays.

Results:

Vector density and behavior: *Anopheles funestus* s.l. (12,758) is the most abundant malaria vector (64% of *Anopheles* species collected). The proportion of *An. gambiae* s.l. (2,111) was 11% of the total *Anopheles* collected. A total of 3,237 *An. tenebrosus* (16%), 692 *An. tchekeidii* (3%), 732 *An. squamosus* (4%), 357 *An. coustani* (2%), 3 *An. rufipes* (0.02%), and 27 *An. ziemanni* (0.1%) were also collected. The mean average of *Anopheles funestus* s.l. was 6 per room per day in the intervention sites in August before IRS campaign and is similar to the density in the sprayed sites in November, one month after IRS (5 *An. funestus* s.l./day/room). The mean indoor resting density of *Anopheles funestus* s.l. dropped from six *Anopheles funestus* s.l. per room per day in August to two in the intervention sites in January three months after IRS. In contrary, in the control sites, the indoor resting density per room per day increased from three *Anopheles funestus* s.l. per room per day before IRS to 11 *Anopheles funestus* s.l. per room per day one month after the IRS campaign and was five *Anopheles funestus* s.l. per room per day in January three months after IRS campaign . five per room per day in January in the control sites..

The mean density of *Anopheles funestus* s.l. in the intervention sites (2.9 *Anopheles funestus* s.l./trap/night) was four times lower than the density in the control sites (12.3/trap/night) in January using the CDC light trap collection method.

The HBR of *Anopheles funestus* s.l. (indoors) was reduced from three bites per person per night during the pre-spray period in August to 0.5 bites per person per night one month after spraying in November in the intervention sites. The drop of the HBR indoors in the intervention sites one month after the IRS campaign might be due to the IRS effect. However, the HBR increased from 0.5 per person per night in November to 4.6 bites per person per night in the

intervention sites in January three months after the spraying. In the control sites, the HBR indoors increased from 10.5 bites per person per night in August to 19 bites per person per night in November and 14.3 bites per person per night in January.

Parity rate: The parity rate is similar in both sprayed and control sites. There is no statistically significant difference between the parity in the sprayed and control sites. The parity is high at most of the sites.

Quality assurance of the 2015 IRS campaign: The WHO cone bioassay performed 24h and one month after spraying showed 100% mortality of the susceptible malaria vectors exposed to the mud and cement sprayed walls.

Insecticide decay rate: Pirimiphos-methyl was effective on both mud and cement in Kasama, Isoka, Katete, and Mwense in February four months after the spraying. The test mortality rate was less than the 80% WHO threshold on the mud and cement sprayed walls in Milenge and Serenje four months after spraying. The mortality rate was above 80% in Isoka and Mwense for both mud and cement sprayed walls and for cement walls in Kasama and Katete, five months after spraying. The residual life of pirimiphos-methyl at all sites was shorter than the expected residual life (six months).

Susceptibility status: *An. funestus* s.l. and *Anopheles gambiae* s.l. were susceptible to pirimiphos-methyl. The resistance intensity assay shows that *Anopheles funestus* s.l. was resistant to all the four pyrethroids tested at all selected sites in Milenge and Samfiya districts. No difference was observed between the sites in the intensity of deltamethrin resistance in the area except in Shitambulli. A difference was observed between the sites in the intensity of permethrin resistance.

1. INTRODUCTION

Malaria is endemic and the transmission is stable with a seasonal peak from November to April in Zambia. (IRS is one of the key malaria elimination strategies of the Zambian National Malaria Control Centre (NMCC)).

In 2014 PMI provided financial and technical support to the NMCC and district health offices for IRS and entomological surveillance activities through the Zambia Integrated Systems Strengthening Program and the PMI AIRS Project.

Entomological surveillance is a key component for IRS programming, providing information on the impact of IRS on vector density and the behavior of malaria vectors in IRS areas compared to non-IRS areas. Entomological activities also help to assess the quality of the IRS operations, the decay rates of the insecticide applied and the vectors susceptibility to insecticides recommended by World Health Organization Pesticides Evaluation Scheme (WHOPES) for use in malaria vector control. The susceptibility data collected are used to select the insecticide for the IRS campaign.

During the 2015 entomological surveillance, the entomological indicators assessed include:

- Malaria vector species composition
- Vector distribution and seasonality (Vector density)
- Vector behavior
- Vector susceptibility to insecticides
- Quality assurance of IRS and decay rate of insecticide applied
- Parity rates

Sporozoite rates and the Entomological Inoculate Rate (EIR) will be provided in the progress report as soon as the enzyme-linked immunosorbent assay (ELISA) circumsporozoite antigens (CSP) results are received from Macha Research Centre.

2. MATERIALS AND METHODS

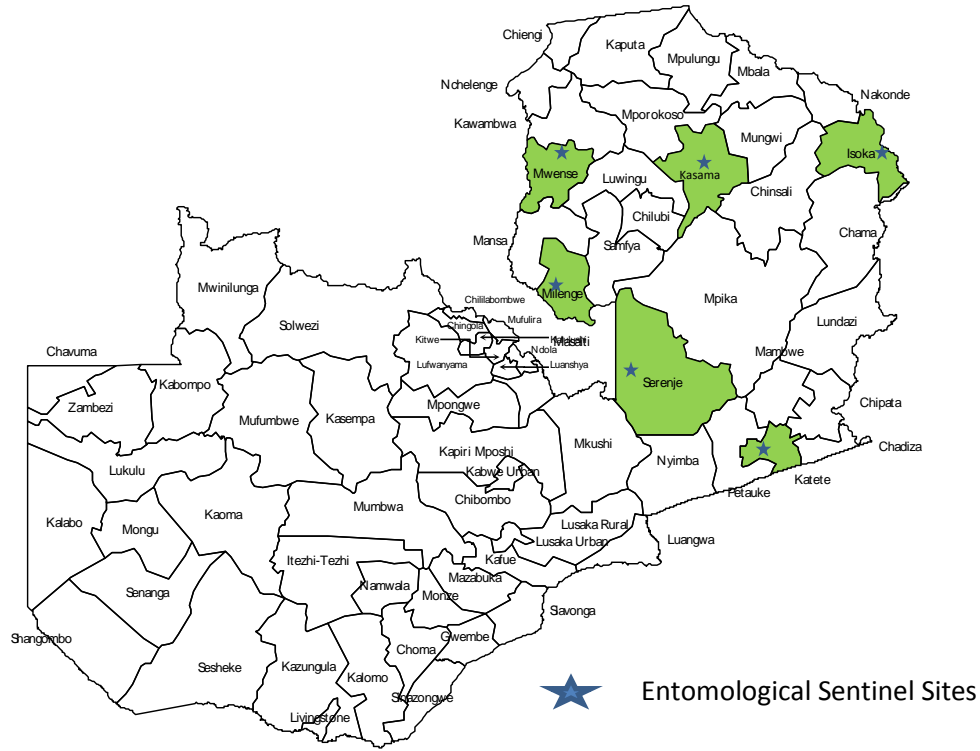
2.1 STUDY SITES

Entomological surveillance was performed in six sentinel districts to assess the 2015 IRS campaign that occurred from September 28 to November 25, 2015. In each district, one sprayed village and another unsprayed village as a control were selected (except in Kasama and Isoka districts where the control sites were sprayed during the 2014 IRS campaign). The sentinel sites covered all five provinces receiving IRS (Table I).

Table 1: Entomological surveillance sentinel sites

Provinces	District	Sites	Spray Status	Percentage of Households Targeted for Spraying
Northern	Kasama	Kalonga	Sprayed	60.5%
Eastern	Katete	Robert	Non-Sprayed (control)	80%
		Mbalani	Sprayed	
Muchinga	Isoka	Nsalamba	Sprayed	87.1%
Central	Serenje	Chbobo	Sprayed	86%
		Chishi	Non-Sprayed (control)	
Luapula	Milenge	Lunga	Sprayed	79%
		Miyambo	Non-Sprayed (control)	
	Mwense	Shibesa	Sprayed	97%
		Chebele	Non-Sprayed (control)	

Figure 1: Geographical locations of entomological sentinel sites in Zambia



2.2 MALARIA VECTOR DENSITY AND BEHAVIOR MONITORING

Adult mosquito collections were done using PSC, CDC light traps, and HLC.

2.2.1 Pyrethrum Spray Catch (PSC)

PSCs were used to sample indoor resting mosquitoes between 4:00 a.m. and 7:00 a.m. in 15 houses for three mornings in each sentinel site. Before the PSC was performed, all occupants were asked to vacate the house without disturbing the resting mosquitoes. The eaves, windows, and other escape routes around the house were sprayed with the pyrethrum mixture (0.025 percent pyrethrum emulsifiable concentrate with 0.1 percent piperonyl-butoxide in kerosene), using a small hand sprayer, followed by spraying of the walls and roof space inside the house. Ten minutes after spraying, all mosquitoes knocked down by the chemical were collected from the white sheets that were placed on the floor before spraying.

2.2.2 CDC light trap collections

CDC light traps were installed inside a total of four houses in each sentinel site for four consecutive nights. The CDC light-trap was suspended in a bedroom 1.5 meters above the floor and about 50 cm from a human sleeping under an insecticide-treated net on the foot side. Traps

were set from 18:00 until 06:00 to ensure that surveillance is conducted during the primary host-seeking periods.

2.2.3 Human Landing Catch (HLC)

The HLC was used to monitor mosquito feeding behavior in all 10 sentinel sites (six sprayed and four control sites). In each targeted village, the collection was done in a total of four houses for four consecutive nights every month. One collector was seated indoors and another seated outdoors from 6 p.m. to 6 a.m. A total of four volunteers per house per night collected from 6 p.m. to 6 a.m. The mosquitoes that tried to bite the volunteer were collected with the mouth aspirator and transferred to paper cups labelled with the hour of collection.

The following parameters were measured from the HLCs:

- Vector feeding behavior (biting time, location of biting)
- Parity
- Biting rate
- Sporozoite rate and EIR will be reported in the progress report as soon as we receive ELISA CSP results from Macha Research Centre.

Community health workers involved in the HLC were provided with chemoprophylaxis with deltaprim.

The mosquitoes collected by PSC, HLC, and CDC light trap were identified. Anophelines were sorted morphologically to species using the Gilles and Coetzee, 1987 identification key. The abdominal status of all female anophelines collected was categorized as unfed, blood-fed, half-gravid and gravid. The anopheline collected were preserved in Eppendorf tubes with silica gel. The preserved malaria vectors (*An. funestus* s.l. and *Anopheles gambiae* s.l.) will be used for polymerase chain reaction (PCR) analysis to identify the sibling species. The sample malaria vectors from the HLC will be analyzed by the ELISA method to look for CSPs and for blood meal determination.

2.3 SUSCEPTIBILITY TESTS

2.3.1 WHO susceptibility test

The CDC Backpack and Prokopack aspirators were used to capture adult indoor-resting mosquitoes. The malaria vectors collected were used in insecticide susceptibility bioassays. Backpack aspiration was performed in as many houses as possible between 04:00 and 07:00 to minimize the chance of mosquitoes leaving the house before the catch is performed. Captured mosquitoes were stored in mosquito cages and were provided access to cotton pads soaked with sugar water to keep them alive.

F1 generation female malaria main vectors aged 2-5 days reared from eggs of field-caught mosquitoes and malaria vectors reared from larvae collected from the field were used for the susceptibility test. The mosquitoes were exposed to diagnostic doses of various insecticides using insecticide-impregnated papers, as described by the WHO guideline. One insecticide from

each class was tested in some of the sentinel sites for *Anopheles funestus* s.l. and *Anopheles gambiae* s.l. A subsample of dead and surviving mosquitoes was preserved and sent to CDC laboratory for the mechanism of the resistance determination. All the resistance study data will be added to the DDMS database.

2.3.2 CDC Resistance Intensity Rapid Diagnostic Test (I-RDT)

AIRS Zambia performed the Rapid Resistance Intensity Diagnostic test (I-RDT) in four districts (Milenge, Samfya, Mwense, and Kawambwa) in Luapula Province to select the sites for an operational research project on resistance intensity.

Adult wild mosquitoes collected through backpack aspiration were used. Prior to the assay, a holding period of one day was observed after the mosquito collection to ensure that any damaged mosquitoes were not used for the test. CDC bottles treated with 1, 2, 5 and 10 times the diagnostic dose of each insecticide plus one control per insecticide were used. Five insecticides were tested with four pyrethroids (deltamethrin, permethrin, lambda-cyhalothrin, alpha-cypermethrin) and PBO.. The mosquitoes were exposed for 30 minutes and the number of mosquitoes knocked down at 0, 15 and 30 minutes was recorded. The mortality was recorded at the diagnostic time (30 minutes). A subsample of dead and surviving mosquitoes was sent to CDC laboratory for the species identification using PCR.

2.4 QUALITY ASSURANCE OF IRS AND INSECTICIDE DECAY RATE

Cone bioassays were used to evaluate the quality of spraying and monitor the decay rate of the insecticide after spraying. The first wall bioassay was conducted 24 hours after spraying in 40 randomly selected houses. Six sprayed houses (three mud, three cement) and two unsprayed (control) houses (one mud and one cement) were used per sentinel site in Mwense, Milenge and Serenje districts. Four sprayed (two mud and two cement) and one control were selected per site in Isoka, Katete, and Kasama districts.

The Kisumu strain of *Anopheles gambiae* s.s. available in the insectary in October was not enough to cover all six sentinel sites for the T0 cone bioassay tests due to the ant invasion and destruction of some mosquitoes in the insectary in July-August. *Anopheles funestus* s.l. was collected using backpack aspirators from Milenge and Mwense districts. Previous surveys showed that *An. funestus* s.l. is most abundant in Milenge and Mwense districts. The WHO susceptibility test was done against pirimiphos-methyl CS (insecticide used for spraying) and showed 100% mortality. This known susceptible *Anopheles funestus* s.l. strain was used for the cone bioassay in Milenge, Mwense, and Serenje districts. The susceptible Kisumu strain (*Anopheles gambiae* s.s.) reared in the insectary at the NMCC was used for the cone bioassay tests in the remaining three districts namely Katete, Isoka, and Kasama. The two most commonly found surface types in the areas, cement and mud walls, were tested.

The cone bioassay tests were done according to the WHO test procedure. The cones were placed on the treated walls at three locations: 0.5m, 1m, and 1.5m above the ground. About 10 females of known susceptible malaria vectors were introduced per cone and exposed for 30 minutes. The number of mosquitoes knocked down after 30 minutes and 60 minutes and dead at the end of 24-hour holding period were recorded.

Subsequent tests were performed at T1 in November, T2 in December, T3 in January, T4 in February, T5 in March, and T6 in April to assess the decay rate of the insecticide applied.

3. RESULTS AND DISCUSSION

3.1 SPECIES COMPOSITION, VECTOR DENSITY, BEHAVIOR AND LONGEVITY

AIRS collected baseline data using indoor CDC light traps, PSC, and HLC in six sprayed sites and four control sites in August to assess the species composition, the vector density and the malaria vectors' behavior before the 2015 IRS campaign. Monthly collections were continued from November to January to assess the impact of the IRS campaign on malaria vectors.

3.1.1 Species composition

A total of 26,416 female mosquitoes were collected using PSC, HLC, and CDC light traps both from spray targeted and control sites, including 6,499 culicine (25%) and 19,917 anopheline (75%). *An. funestus* s.l. (12,758) was the most prevalent species (64%). A total of 2,111 *Anopheles gambiae* s.l. were caught (11%). The other anopheline species found were 3,237 *An. tenebrosus* (16%), 692 *An. tchekeidii* (3%), 732 *An. squamosus* (4%), 357 *An. coustani* (2%), 3 *An. rufipes* (0.0006%) and 27 *An. ziemanni* (0.0014%). Tables A-H in the Annex show the number of each species collected per sites and per month.

Figure 2: Species composition in the targeted sprayed sentinel sites (Aug 2015 - Jan 2016)

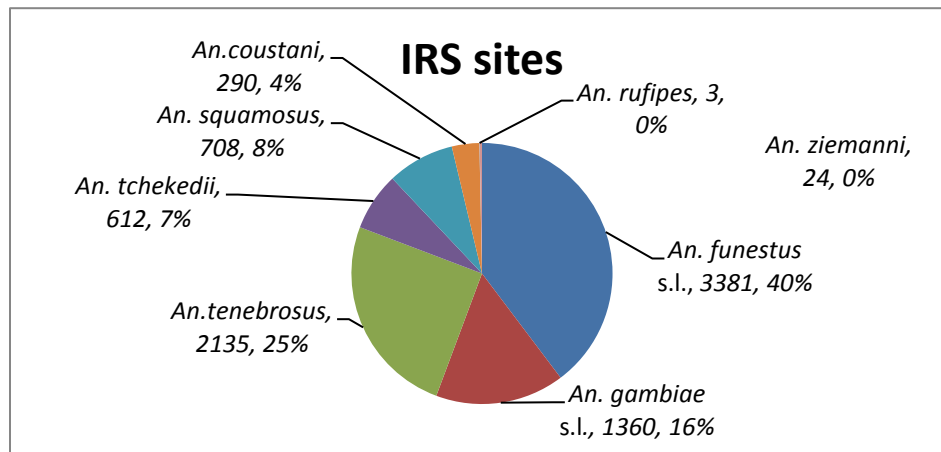
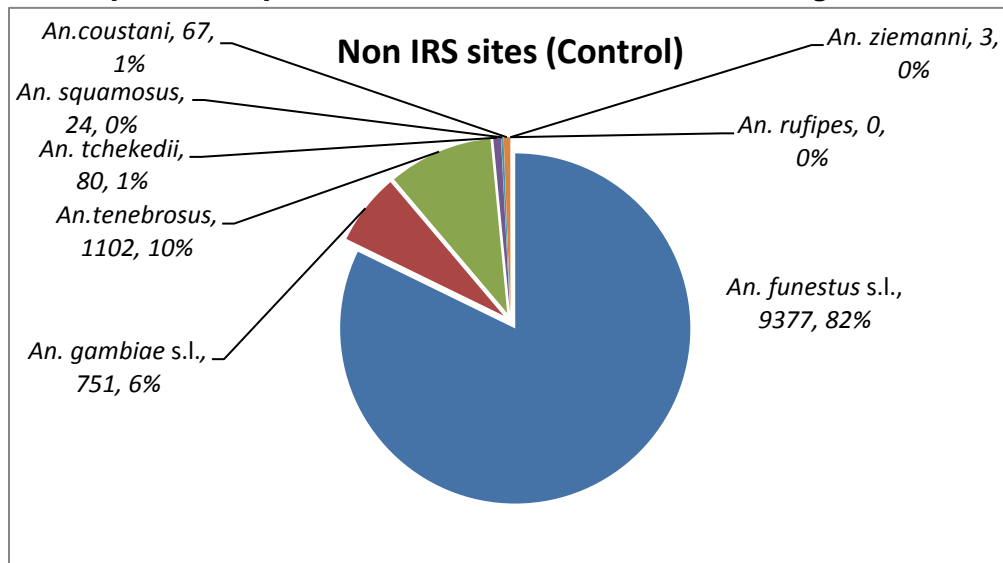


Figure 3 : Species composition in the control sentinel sites (Aug 2015 - Jan 2016)



3.2 VECTOR DENSITY, BEHAVIOR, AND LONGEVITY

3.2.1 Pyrethrum Spray Catch

As indicated in Tables 2, 3, 4, and 5, the average density of *Anopheles funestus* s.l. collected by PSC both from the intervention and control sites was low in August prior to the IRS campaign except in Milenge and Mwense. The highest density was recorded in Milenge where 28 *Anopheles funestus* s.l. were collected per room per day in Lunga (sprayed site) and nine *Anopheles funestus* s.l. per room per day in Miyambo (control site) in August. The average density in Lunga (7.9 *Anopheles funestus* s.l./room/day) was four times lower in January, three months after IRS campaign as compared to the density recorded in August. The average density of *An. funestus* s.l. per room per day in Shibesa dropped from 7.25 before IRS to 2.75 in January, three months after IRS campaign. Less than one *Anopheles funestus* s.l. was collected in Kalonga and Nsalamba during the study period. No *Anopheles funestus* s.l. was found in Katete District.

Overall, The mean average of *Anopheles funestus* s.l. was 6 per room per day in the intervention sites in August before IRS campaign and is similar to the density in the sprayed sites in November , one month after IRS (5 *An.funestus* s.l./day/room). The mean indoor resting density of *Anopheles funestus* s.l. dropped from six *Anopheles funestus* s.l. per room per day in August to two in January three months after IRS. In contrary, in the control sites, the indoor resting density per room per day increased, from three *Anopheles funestus* s.l. per room per day before IRS in August to 11 *Anopheles funestus* s.l. per room per day one month after the IRS campaign and was five *Anopheles funestus* s.l. per room per day three months after spraying

The proportion of half and full gravid *Anopheles funestus* s.l. was similar and high in both sprayed and control sites (32% in sprayed sites and 31% in control sites), after the IRS campaign. More than half of the *Anopheles funestus* s.l. collected inside the houses (59.8% in the

intervention sites and 54% in the control) by PSC was found fed. The blood meal analysis will confirm if the vector-man contact is maintained even after IRS.

Table 2 : Blood digestion stage and average density of *An. funestus* s.l. collected by PSC per room in Serenje District in August and from November 2015 to January 2016

Months	Number of rooms	<i>An. funestus</i> s.l. Collected	Physiological age				Average density per room
			UF	F	HG	G	
Chibobo (Sprayed site)							
August (Baseline)	8	0	0	0	0	0	0
November	15	40	4	6	15	15	2.7
December	15	27	2	19	1	5	1.8
January	15	34	3	24	5	2	2.3
Chishi (Control site)							
August (Baseline)	8	1	1	0	0	0	0.1
November	15	28	3	16	8	1	1.9
December	15	93	9	50	19	15	6.2
January	15	108	11	84	9	4	7.2

Table 3 : Blood digestion stage and average density of *An. funestus* s.l. collected by PSC per room in Milenge District in August and from November 2015 to January 2016

Months	Number of rooms	<i>An. funestus</i> s.l. Collected	Physiological age				Average density per room
			UF	F	HG	G	
Lunga (Sprayed site)							
August (Baseline)	8	224	83	131	0	10	28
November	15	393	27	234	93	39	26.2
December	15	65	4	35	5	21	4.3
January	15	119	9	90	11	9	7.9
Miyambo (Control site)							
August (Baseline)	8	74	4	70	0	0	9.25
November	15	568	87	284	98	99	37.9
December	15	243	40	120	35	48	16.2
January	15	155	36	109	1	9	10.3

Table 4 : Blood digestion stage and average density of *An. funestus* s.l. collected by PSC per room in Mwense District in August and from November 2015 to January 2016

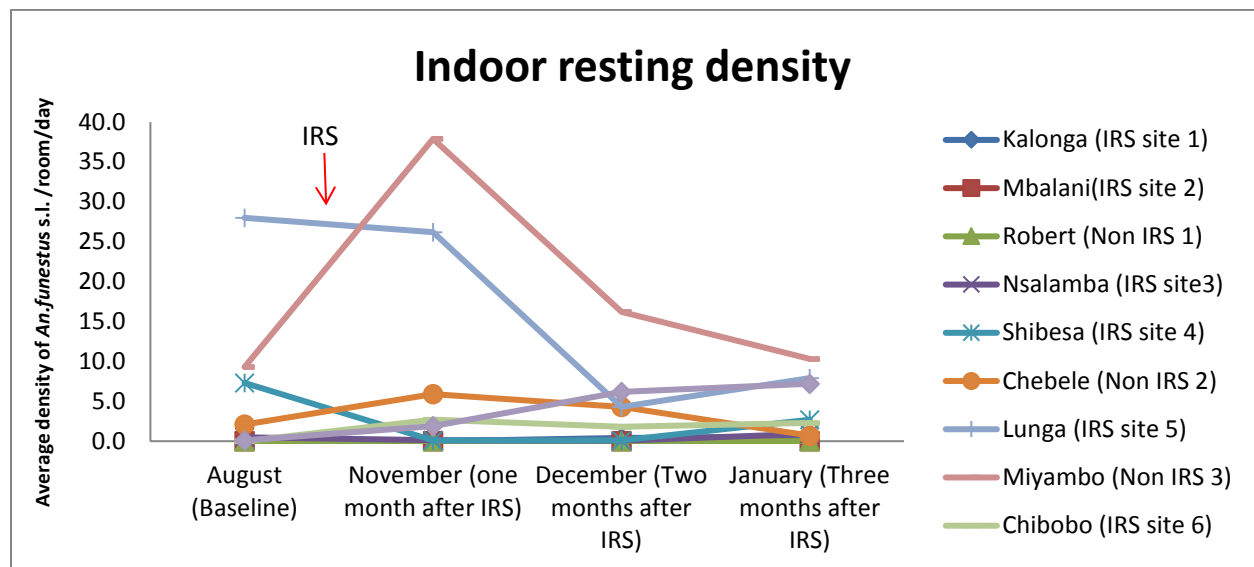
Months	Number of rooms	<i>An. funestus</i> s.l. Collected	Physiological age				Average density per room
			UF	F	HG	G	
Shibesa (Sprayed site)							
August (Baseline)	8	58	5	53	0	0	7.25
November	15	1	0	1	0	0	0.1
December	15	2	0	2	0	0	0.1
January	15	41	3	29	2	7	2.7
Chebele (Control sites)							
August (Baseline)	8	17	4	13	0	0	2.1
November	15	89	7	36	30	16	5.9
December	15	64	9	27	15	13	4.3
January	15	11	3	8	0	0	0.7

Table 5 : Blood digestion stage and average density of *An. funestus* s.l. collected by PSC per room in Kasama and Isoka districts in August and from November 2015 to January 2016

Months	Number of rooms	<i>An. funestus</i> s.l. Collected	Physiological age				Average density per room
			UF	F	HG	G	
Kalonga (Sprayed site) Kasama district							
August (Baseline)	8	0	0	0	0	0	0
November	15	0	0	0	0	0	0
December	15	6	3	3	0	0	0.4
January	15	9	0	2	4	3	0.6
Nsalamba (Sprayed site) Isoka district							
August (Baseline)	8	4	2	2	0	0	0.5

November	15	2	2	0	0	0	0.13
December	15	1	1	0	0	0	0.07
January	15	14	3	6	1	4	0.93

Figure 4: Indoor resting density of *Anopheles funestus* s.l. per room per night in August (baseline) and one, two and three months after IRS



A total of 128 *Anopheles gambiae* s.l. were collected by PSC from the sprayed and control sites. A total of 79 *Anopheles gambiae* s.l. were collected in the sprayed sites and 49 in the control sites. *Anopheles gambiae* s.l. was not found in the rooms in Serenje and Katete districts. The average density of *Anopheles gambiae* s.l. was low, less than one *Anopheles gambiae* s.l. in Milenge and Kasama districts. The highest density was recorded in Mwense District where 3.4 *Anopheles gambiae* s.l. were found per room per day in the sprayed site and 2.6 in the control. Figures 5, 6, and 7 show the average density per room per day in Mwense, Milenge, Isoka, and Kasama districts.

Figure 5: Average density of *Anopheles gambiae* s.l. per room per day in Mwense District (Aug 2015 - Jan 2016)

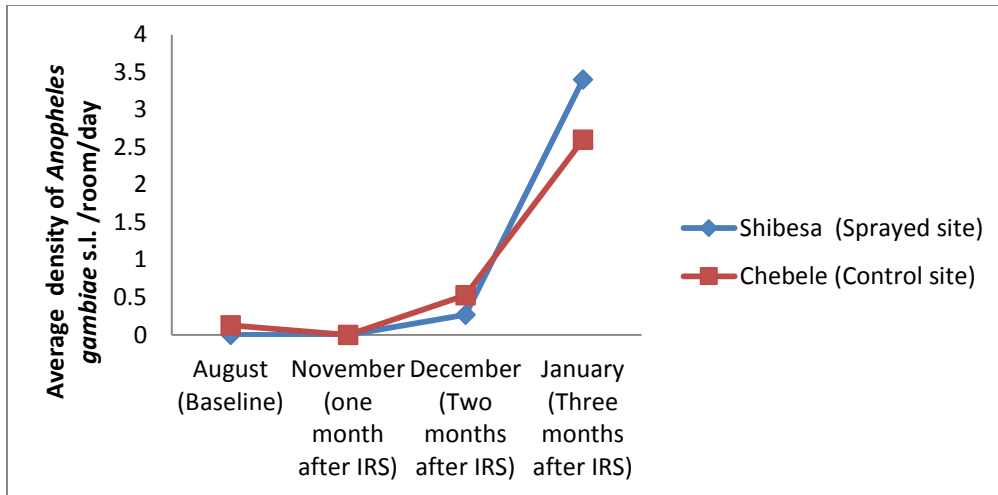


Figure 6: Average density of *Anopheles gambiae* s.l. per room per day in Milenge District (Aug 2015 - Jan 2016)

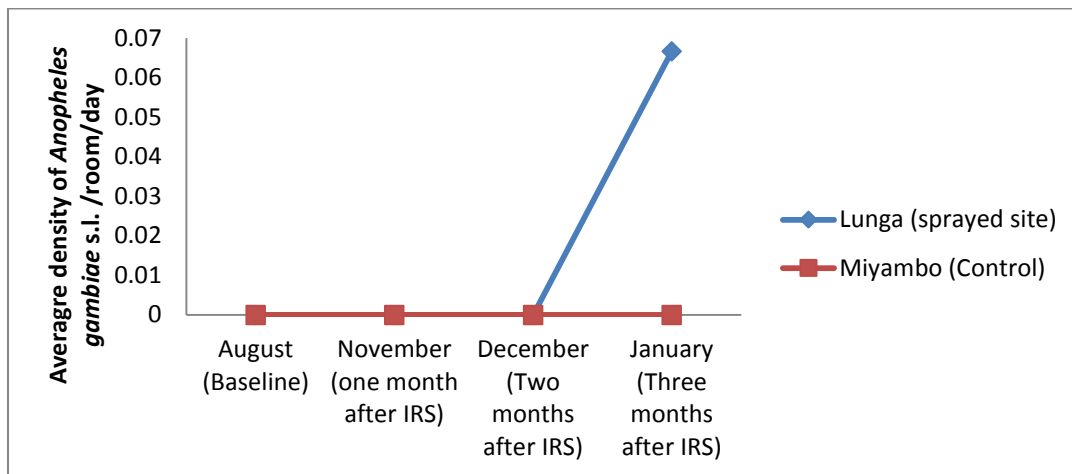
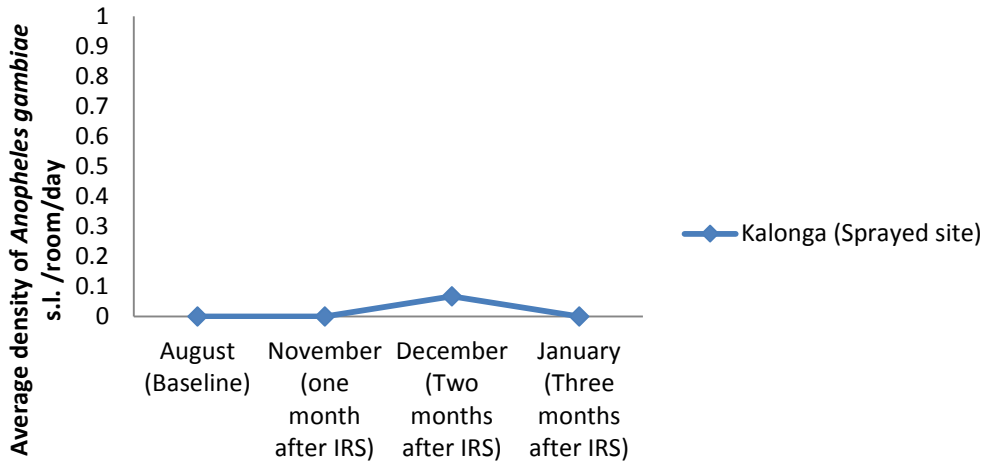


Figure 7: Average density of *Anopheles gambiae* s.l. per room per day in Kasama and Isoka Districts (Aug 2015 - Jan 2016)

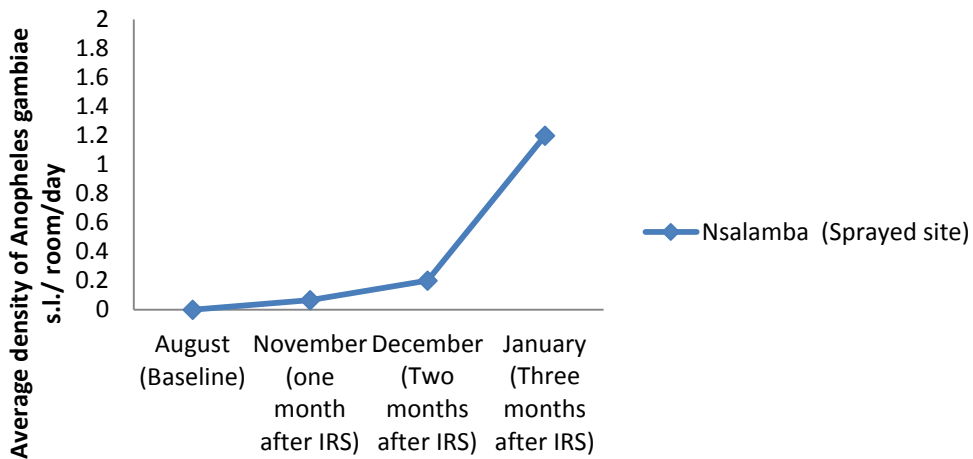
a) Kasama district

Indoor resting density in Kasama district



b) Isoka district

Indoor resting density in Isoka



3.2.2 CDC light trap collection

The average density of *An. funestus* s.l. per trap per night in the intervention sites dropped from five to 0.7 one month after IRS campaign. The density of *Anopheles funestus* s.l. per trap per night in the sprayed sites was 2.9 three months after the IRS campaign in January in the intervention sites. In contrary, the density of *Anopheles funestus* s.l. increased from seven in August to 12.2 per trap per night one month after IRS and was still high (12.3 per trap per night) three months in January in the control sites. The mean density of *Anopheles funestus* s.l. in the intervention site (2.9 *Anopheles funestus* s.l. /trap/night) was four times lower than the density in the control sites (12.3/trap/night) in January.

The average density of *An. funestus* s.l. in Miyambo (24 *Anopheles funestus* s.l./trap/night) in Milenge District was two times higher than the density recorded in Lunga, the intervention site (13.3 *Anopheles funestus* s.l. /trap/night) in August before IRS. The density decreased from 13 *Anopheles funestus* s.l./ trap/night to 4.1 two months after spraying, but increased to 13 *Anopheles funestus* s.l. /trap/night in Lunga three months after IRS in January. However, the density recorded in Miyambo, the control site, was three times higher than the density of *Anopheles funestus* s.l. in Lunga in January 2016.

Anopheles gambiae s.l. was not collected in Katete District by CDC light trap during the study period. Less than one *Anopheles gambiae* s.l. was caught per trap per night in the sprayed sites except in January when the average density per trap per night was between zero in Chibobo in Serenje District and 14.4 in Nsalamba in Isoka District.

Figure 8: Average density of *Anopheles funestus* s.l. per trap per night in Katete District (Aug 2015 - Jan 2016)

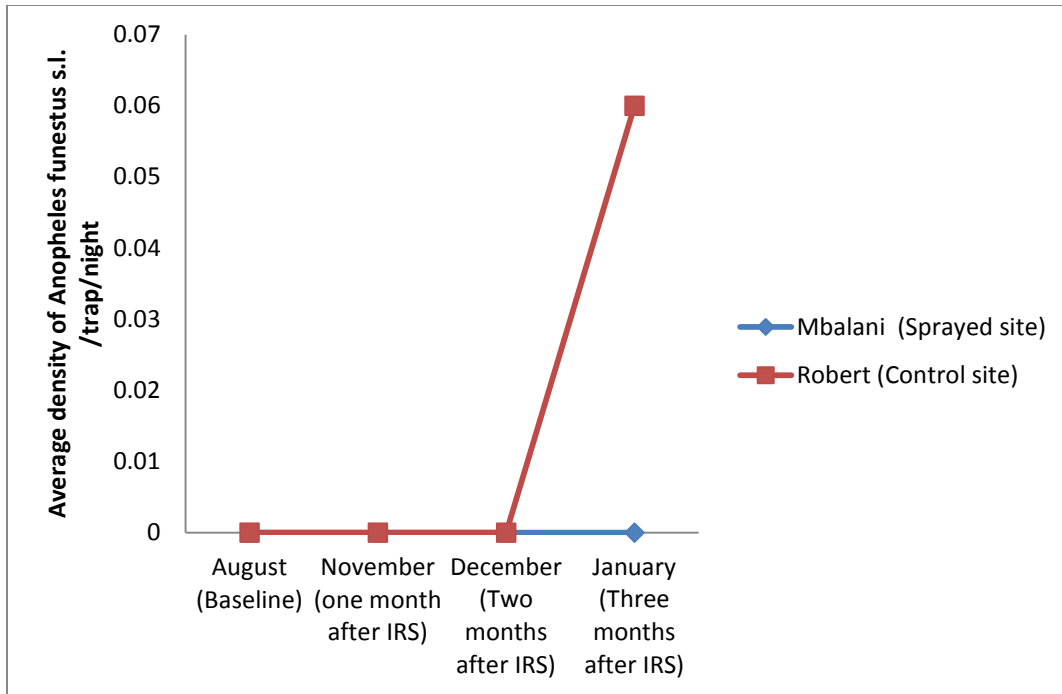


Figure 9: Average density of *Anopheles funestus* s.l. per trap per night in Mwense District (Aug 2015 - Jan 2016)

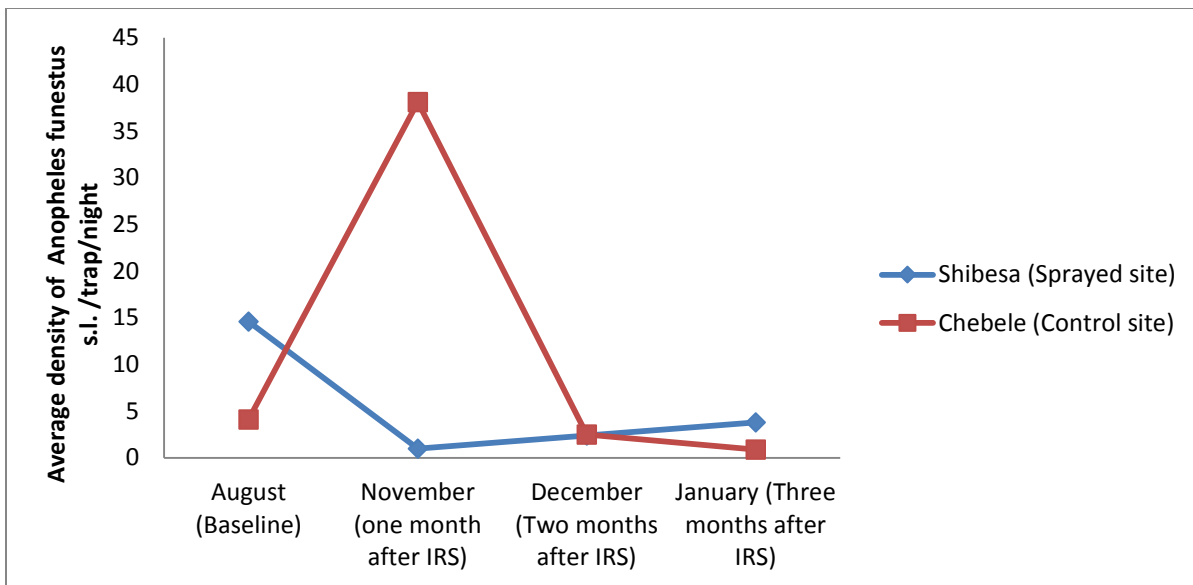


Figure 10: Average density of *Anopheles gambiae* s.l. per trap per night in Mwense District (Aug 2015 - Jan 2016)

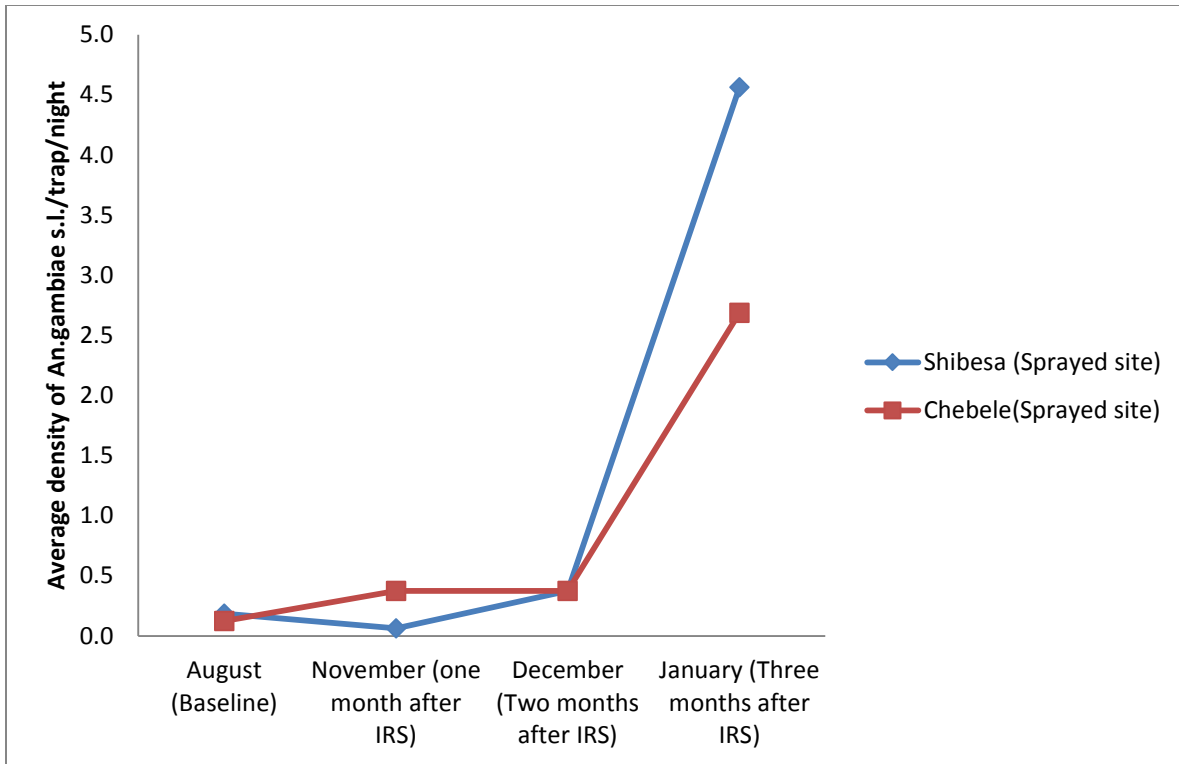


Figure 11 : Average density of *Anopheles funestus* s.l. per trap per night in Milenge District (Aug 2015 - Jan 2016)

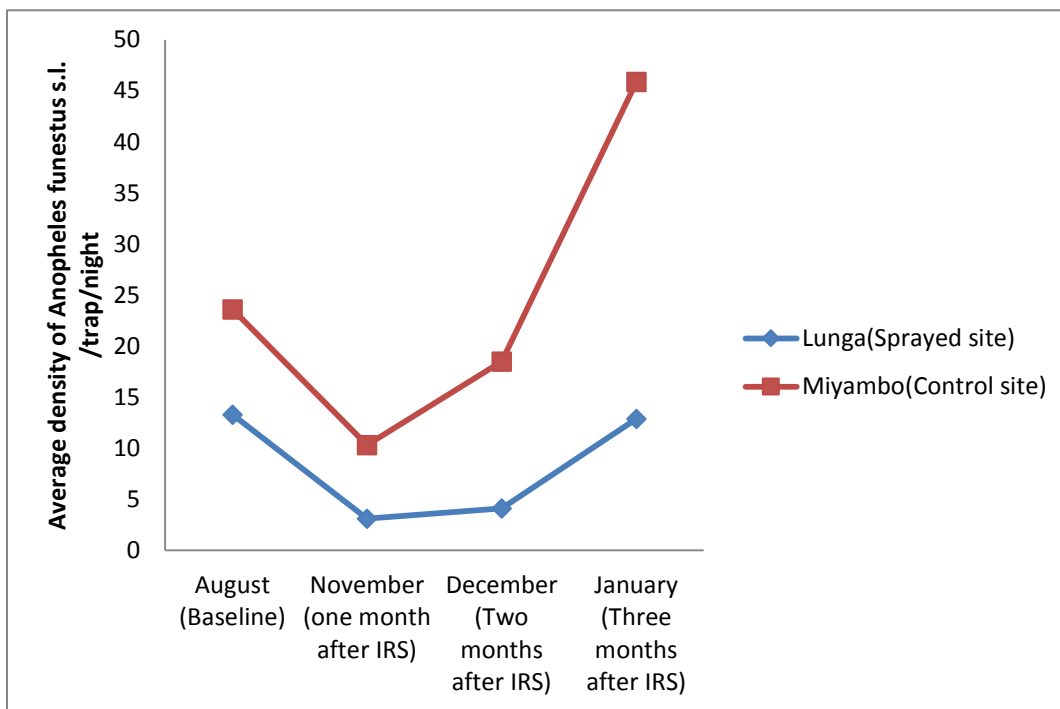


Figure 12: Average density of *Anopheles gambiae* s.l. per trap per night in Milenge District (Aug 2015 - Jan 2016)

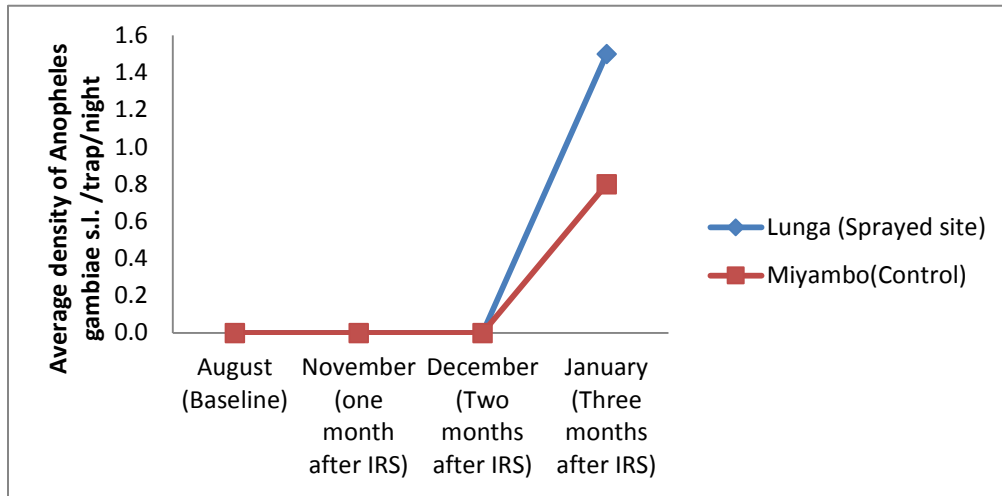


Figure 13: Average density of *Anopheles funestus* s.l. per trap per night in Serenje District (Aug 2015 - Jan 2016)

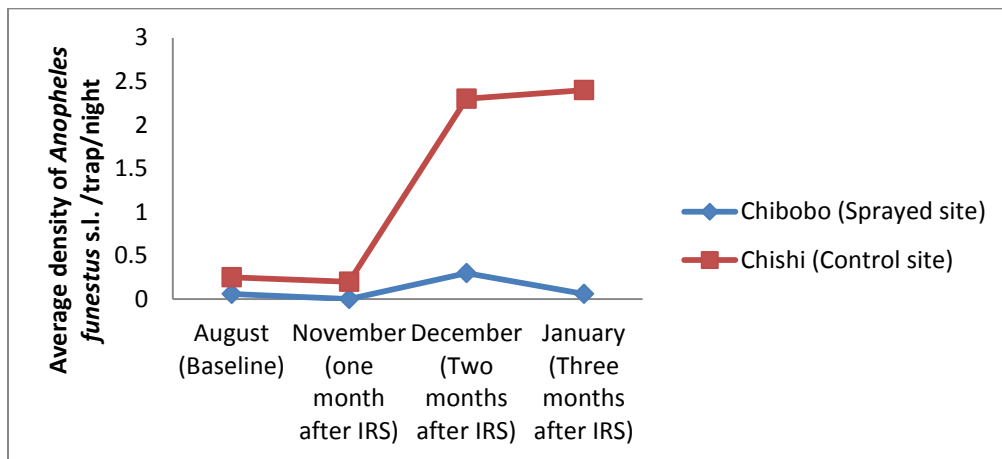
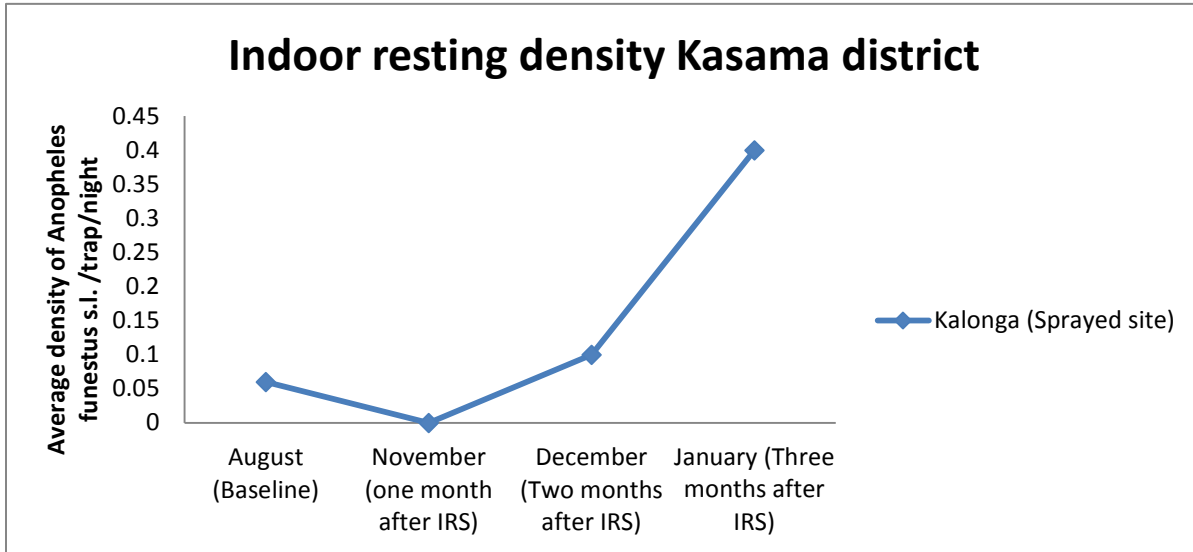


Figure 14: Average density of *Anopheles funestus* s.l. per trap per night in Kasama and Isoka districts (Aug 2015 - Jan 2016)

a) Kasama district



b) Isoka district

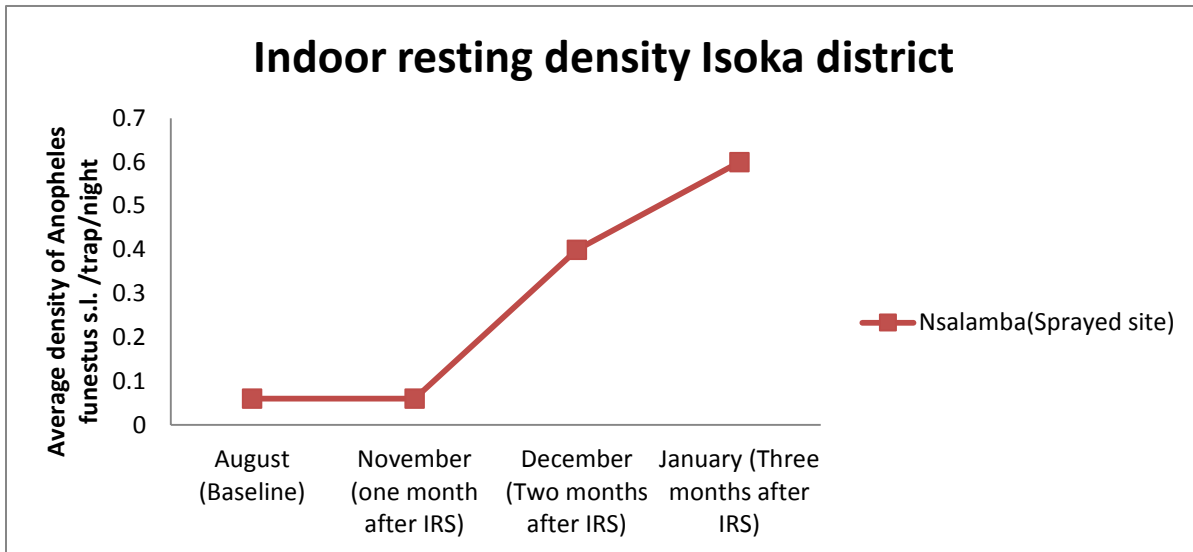
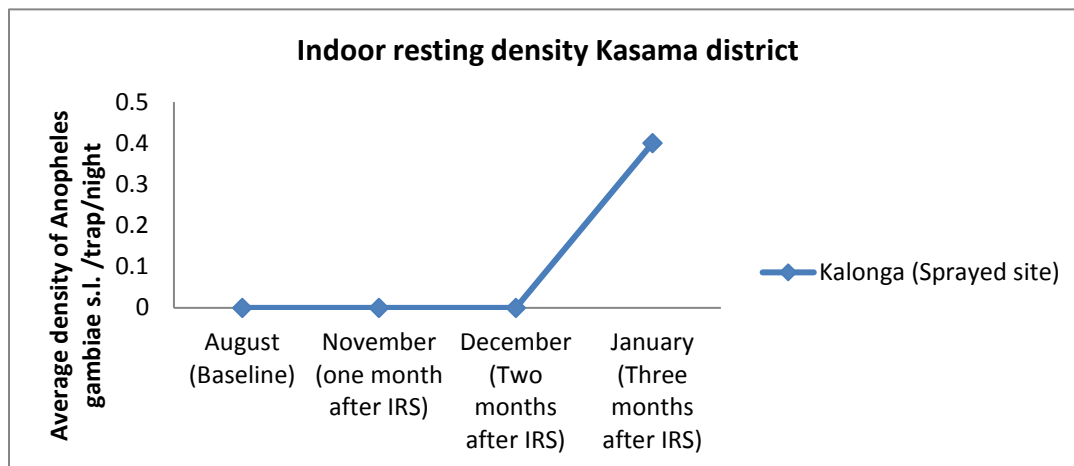
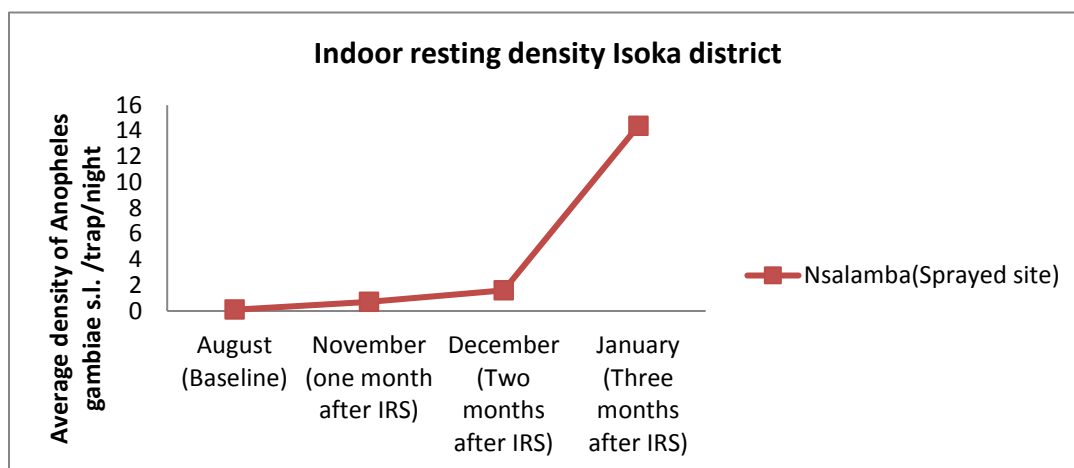


Figure 15: Average density of *Anopheles gambiae* s.l. per trap per night in Kasama and Isoka Districts from August 2015 to January 2016

a) Kasama district



b) Isoka district



3.2.3 HLC Collection

During the collection period, 13,025 female *Anopheles* were collected indoors and outdoors from the sprayed and control sites. A total of 6,965 *Anopheles funestus* s.l. and 1,532 *Anopheles gambiae* s.l. were collected. *An. tenebrosus* (2,837), *An. tchekeidii* (616), *An. squamosus* (692), *An. rufipes* (3), *An. coustani* (353) and *An. ziemanni* (27) were also collected biting indoors and outdoors. Figure 16 gives the composition of *Anopheles* species collected by HLC.

Figure 16: *Anopheles* species composition of the indoor HLC (Aug 2015 to Jan 2016)

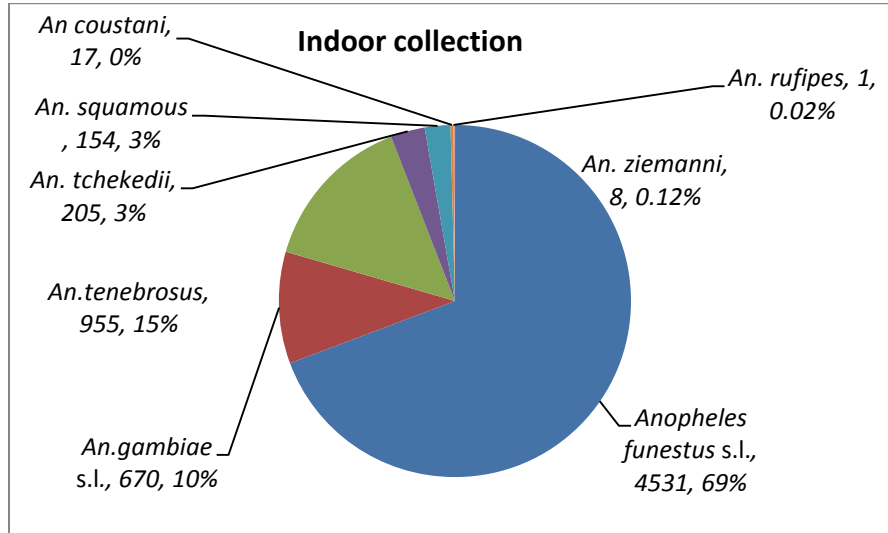
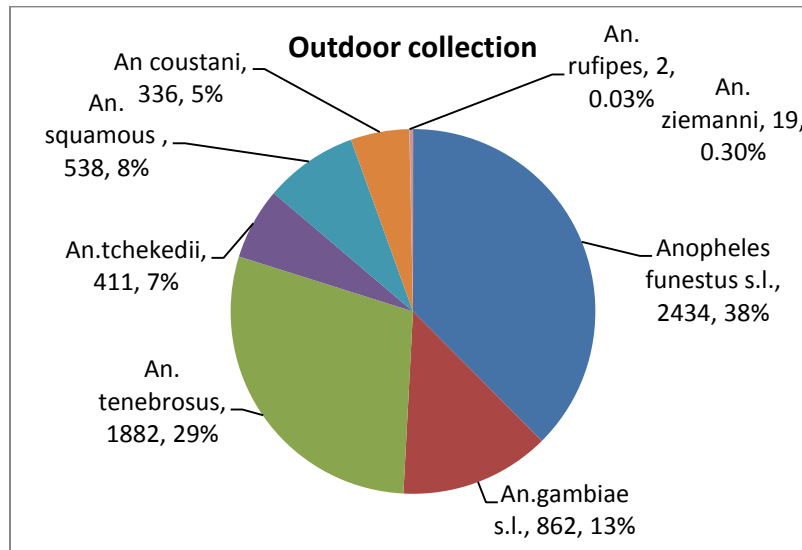


Figure 17: *Anopheles* species composition of the outdoor HLC (Aug 2015 to Jan 2016)



Human biting rate of *Anopheles funestus* s.l.

The mean biting rate of *Anopheles funestus* s.l. was 10.5 per person per night inside the house and 4.3 per person per night outdoors in the control sites versus three per person per night inside and 1.3 per person per night outdoors in the spray targeted site in August.

The human biting rate of *Anopheles funestus* s.l. was reduced from three bites per person per night during the pre-spray period to 0.5 bites per person per night one month after spraying in the intervention sites. The drop of the human biting rate in the intervention sites one month after IRS campaign might be due to the IRS effect. However, the biting rate increased from 0.5 per person per night in November to 4.6 bites per person per night in the intervention sites in January three months after the spraying. In the control sites, the HBR indoors increased from 10.5 bites per person per night in August to 19 bites per person per night in November and 14.3 bites per person per night in January.

In Katete District during the pre-spray period and three months after spraying, no malaria vectors were collected from Mbalani, the sprayed site. In the control site the HBR indoors was low. Each person received only two *Anopheles funestus* s.l. bites per month inside house (0.0625 bites/night) in Robert (control site) in December and January versus no bites per month in the intervention site.

The HBR indoors in Shibesa (sprayed site) in Mwense District was 0.25 *Anopheles funestus* s.l. bites per person per night in November one month after spraying —240 times lower than the HBR in the control sites of Chebele (60 *Anopheles funestus* s.l. bites/person/night). However, the HBR inside houses increased from 0.25 to 3 bites per person per night in January and is similar to the HBR recorded indoors in the control site in January (2.75 bites/person/night).

The HBR indoors in Lunga (sprayed) in Milenge was 14 bites per person per night during the pre-spray period and dropped to 2.3 bites per person per night one month after IRS. The rate increased to 20 bites per person per night in January three months after spraying. The HBR indoors in Miyambo, the control site, increased from 16.25 bites/person/night in November to 52 bites/person/night.

Table 6 provides the HBR of *Anopheles funestus* s.l. from August to January in the control and sprayed sites. Tables 7, 8, 9, 10, and 11 show the HBR per district.

Table 6: *Anopheles funestus* s.l. HBR in sprayed and control sites in August and from November 2015 to January 2016

		August	November	December	January
Total Intervention sites					
Indoor	Total <i>Anopheles funestus</i> s.l.	228	44	160	441
	nb collectors	20	24	24	24
	nb nights	4	4	4	4
	HBR/night	2.9	0.5	1.7	4.6
Outdoor	Total <i>Anopheles funestus</i> s.l.	105	32	134	282
	nb collectors	20	24	24	24
	nb nights	4	4	4	4
	HBR/night	1.3	0.3	1.4	3

		Total Control sites			
Indoor	Total <i>Anopheles funestus</i> s.l.	503	1221	1021	913
	nb collectors	12	16	16	16
	nb nights	4	4	4	4
	HBR/night	10.5	19.1	16	14.3
Outdoor	Total <i>Anopheles funestus</i> s.l.	205	602	545	529
	nb collectors	12	16	16	16
	nb nights	4	4	4	4
	HBR/night	4.3	9.4	8.5	8.3

Table 7: *Anopheles funestus* s.l. HBR in Katete District in August and from November 2015 to January 2016

		August	November	December	January
Mbalani (Intervention site)					
Indoor	Total <i>Anopheles funestus</i> s.l.	0	0	0	0
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0	0
Outdoor	Total <i>Anopheles funestus</i> s.l.	0	0	0	0
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0	0
Robert (Control)					
Indoor	Total <i>Anopheles funestus</i> s.l.	1	0	1	1
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0.0625	0	0.0625	0.0625
Outdoor	Total <i>Anopheles funestus</i> s.l.	0	0	1	0
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0.0625	0

Table 8: *Anopheles funestus* s.l. HBR in Mwenze District in August and from November 2015 to January 2016

		August	November	December	January
Shibesa (Intervention site)					
Indoor	Total <i>Anopheles funestus</i> s.l.		4	18	49
	nb collectors		4	4	4
	nb nights		4	4	4
	HBR/night		0.25	1.125	3.0625
Outdoor	Total <i>Anopheles funestus</i> s.l.		1	31	40
	nb collectors		4	4	4
	nb nights		4	4	4
	HBR/night		0.06	1.93	2.5
Chebele (Control)					
Indoor	Total <i>Anopheles funestus</i> s.l.		960	179	44
	nb collectors		4	4	4
	nb nights		4	4	4
	HBR/night		60	11.1875	2.75
Outdoor	Total <i>Anopheles funestus</i> s.l.		510	100	36
	nb collectors		4	4	4
	nb nights		4	4	4
	HBR/night		31.875	6.25	2.25

Table 9 : *Anopheles funestus* s.l. HBR in Milenge District in August and from November 2015 to January 2016

		August	November	December	January
Lunga (Intervention site)					
Indoor	Total <i>Anopheles funestus</i> s.l.	220	36	126	327
	nb collectors s	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	13.75	2.25	7.875	20.4375
Outdoor	Total <i>Anopheles funestus</i> s.l.	93	17	79	176
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	5.8125	1.0625	4.9375	11
Miyambo (Control)					
Indoor	Total <i>Anopheles funestus</i> s.l.	497	260	759	828

	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	31.0625	16.25	47.4375	51.75
Outdoor	Total <i>Anopheles funestus</i> s.l.	202	91	397	483
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	12.625	5.6875	24.8125	30.1875

Table 10: *Anopheles funestus* s.l. HBR in Mwense District in August and from November 2015 to January 2016

		August	November	December	January
Chibobo (Intervention site)					
Indoor	Total <i>Anopheles funestus</i> s.l.	0	3	6	12
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0.1875	0.375	0.75
Outdoor	Total <i>Anopheles funestus</i> s.l.	0	0	15	10
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0.9375	0.625
Chishi (control)					
Indoor	Total <i>Anopheles funestus</i> s.l.	5	1	82	40
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0.3125	0.0625	5.125	2.5
Outdoor	Total <i>Anopheles funestus</i> s.l.	3	1	47	10
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0.1875	0.0625	2.9375	0.625

Table 11: *Anopheles funestus* s.l. biting rate in Kasama and Isoka districts in August and from November 2015 to January 2016

		August	November	December	January
Kalonga (Intervention site)					

Indoor	Total <i>Anopheles funestus</i> s.l.	6	0	8	31
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0.375	0	0.5	1.9375
Outdoor	Total <i>Anopheles funestus</i> s.l.	1	1	2	16
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0.0625	0.0625	0.125	1
Nsalamba (Intervention site)					
Indoor	Total <i>Anopheles funestus</i> s.l.	2	1	2	22
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0.125	0.0625	0.125	1.375
Outdoor	Total <i>Anopheles funestus</i> s.l.	11	13	7	40
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0.6875	0.8125	0.4375	2.5

***Anopheles funestus* s.l. biting time**

Anopheles funestus s.l. started blood feeding indoors early in the evening in Lunga in Milenge in August. The peak of the biting rate was recorded between 0:00-1:00 a.m. After the IRS campaign, two peaks were observed, one between 1:00 -2:00 am and the second between 5:00-6:00 am.

Anopheles funestus s.l. started biting indoors between 7:00-8:00 pm. The biting peak was observed between 0:00 and 1:00 am after IRS in Shibesa in Mwense.

The peak biting of *Anopheles funestus* s.l. was recorded between 3:00 and 4:00 am in Chibobo in Serenje District, between 4:00-5:00 am in Kalonga in Kasama, and between 3:00 and 4:00 am in Nsalamba in Isoka District.

Figure 18: *Anopheles funestus* s.l. feeding time in Lunga sprayed site in Milenge District, in August 2015 before IRS and from November to January 2015 after IRS

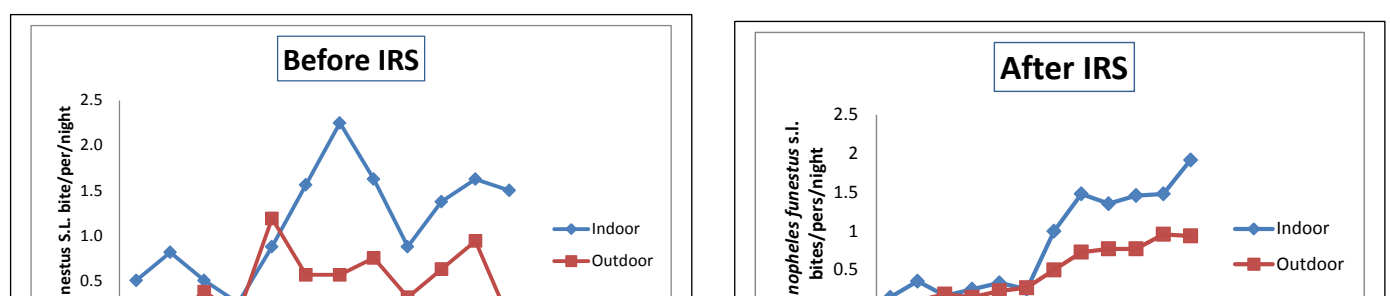


Figure 19: *Anopheles funestus* s.l. feeding time in Shibesa sprayed site in Mwense District from November to January 2015 after IRS

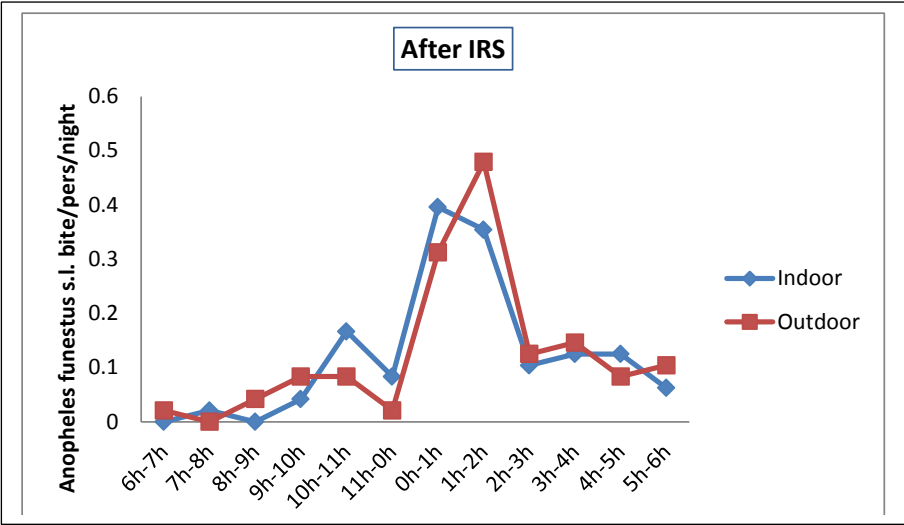
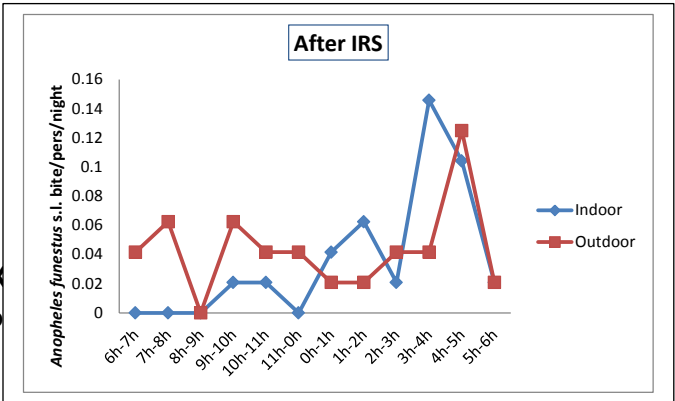
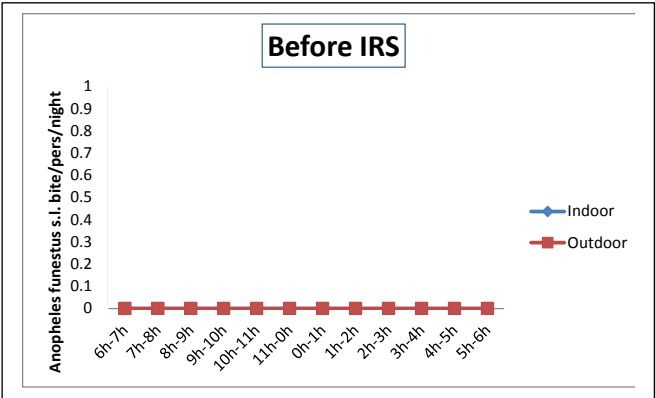


Figure 20: *Anopheles funestus* s.l. feeding time in Chibobo sprayed site in Serenje district in August 2015 before IRS and from November to January 2015 after IRS



in K No

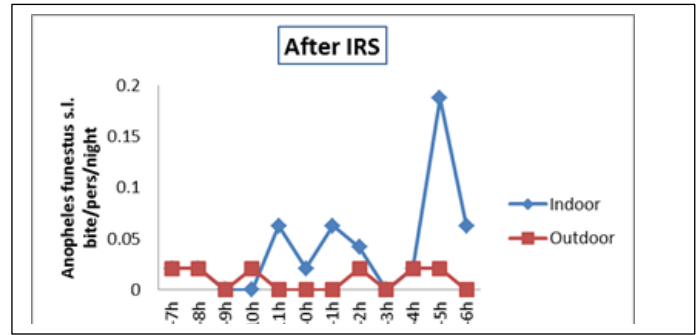
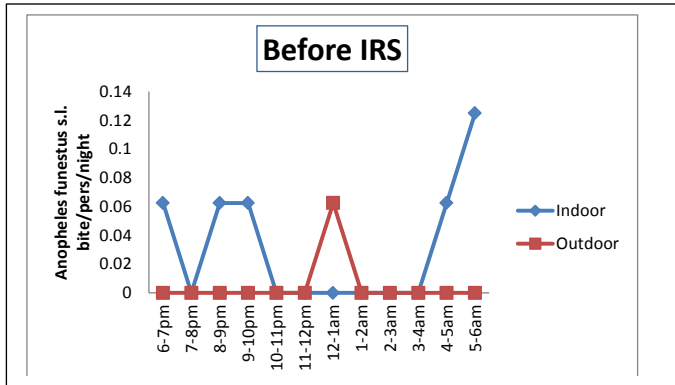
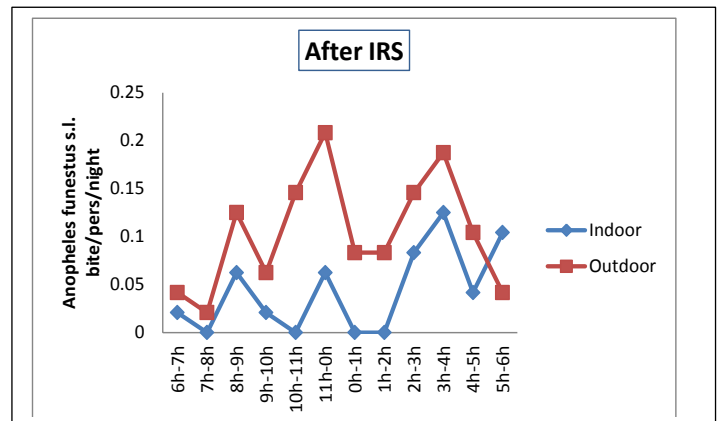
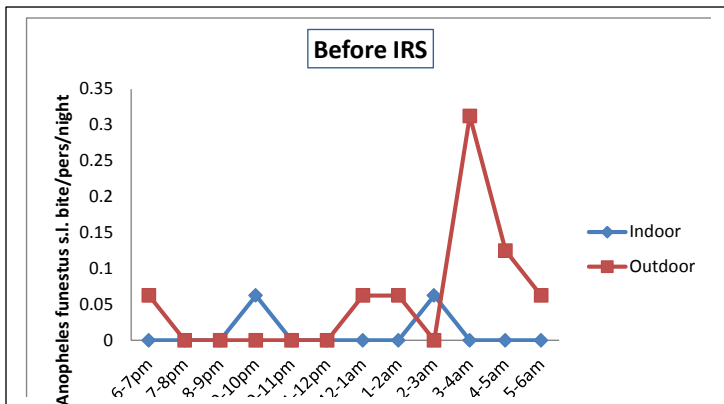


Figure 22: *Anopheles funestus* s.l. feeding time in Nsalamba sprayed site in Isoka District in

August 2015 before and from November to January after IRS



3.3 ANOPHELES GAMBIAE S.L. BITING RATE

Anopheles gambiae s.l. was not found biting in August one month before IRS except in Nsalamba in Isoka where it was found biting both indoors and outdoors. The biting rate in Shibesa was five *Anopheles gambiae* s.l. bites per person per night inside house in January three months after spraying. The highest biting rate of *Anopheles gambiae* s.l. (14 bites/person/night) inside was recorded in Nsalamba in Isoka District in January. *Anopheles gambiae* s.l. was not collected in Serenje during the study period. Tables 11, 12, 13, and 14 provide the HBR per district from August to January.

Table 12: *Anopheles gambiae* s.l. biting rate in Milenge District in August and from November 2015 to January 2016

		August	November	December	January
Lunga (Intervention site)					
Indoor	Total <i>Anopheles gambiae</i> s.l.	0	0	0	11
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0	0.6875
Outdoor	Total <i>Anopheles gambiae</i> s.l.	0	0	1	8
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0.0625	0.5
Miyambo (Control)					
Indoor	Total <i>Anopheles gambiae</i> s.l.	0	0	2	7
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0.125	0.4375
Outdoor	Total <i>Anopheles gambiae</i> s.l.	0	0	0	11
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0	0.6875

Table 13: *Anopheles gambiae* s.l. biting rate in Mwense District in August and from November 2015 to January 2016

		August	November	December	January
Shibesa (Intervention site)					
Indoor	Total <i>Anopheles gambiae</i> s.l.		5	27	80
	nb collectors		4	4	4
	nb nights		4	4	4
	HBR/night		0.3125	1.6875	5
Outdoor	Total <i>Anopheles gambiae</i> s.l.		2	36	70
	nb collectors		4	4	4
	nb nights		4	4	4
	HBR/night		0.125	2.25	4.375
Chebele (Control)					
Indoor	Total <i>Anopheles gambiae</i> s.l.		70	52	151
	nb collectors		4	4	4
	nb nights		4	4	4
	HBR/night		4.375	3.25	9.4375
Outdoor	Total <i>Anopheles gambiae</i> s.l.		100	68	164
	nb collectors		4	4	4
	nb nights		4	4	4
	HBR/night		6.25	4.25	10.25

Table 14: *Anopheles gambiae* s.l. biting rate in Serenje District in August and from November 2015 to January 2016

		August	November	December	January
Chibobo (Intervention site)					
Indoor	Total <i>Anopheles gambiae</i> s.l.	0	0	0	0
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0	0

Outdoor	Total <i>Anopheles gambiae</i> s.l.	0	0	0	0
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0	0
Chishi (Control)					
Indoor	Total <i>Anopheles gambiae</i> s.l.	0	0	0	0
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0	0
Outdoor	Total <i>Anopheles gambiae</i> s.l.	0	0	0	0
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0	0

Table 15: *Anopheles gambiae* s.l. biting rate in Kasama and Isoka districts in August and from November 2015 to January 2016

		August	November	December	January
Kalonga (Intervention site)					
Indoor	Total <i>Anopheles gambiae</i> s.l.	0	0	1	6
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0.0625	0.375
Outdoor	Total <i>Anopheles gambiae</i> s.l.	0	0	3	3
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0	0	0.1875	0.1875
Nsalamba (Intervention site)					
Indoor	Total <i>Anopheles gambiae</i> s.l.	5	15	8	222
	nb collectors	4	4	4	4
	nb nights	4	4	4	4
	HBR/night	0.3125	0.9375	0.5	13.875
Outdoor	Total <i>Anopheles gambiae</i> s.l.	2	18	44	332
	nb collectors	4	4	4	4
	nb nights	4	4	4	4

3.4 ANOPHELES GAMBIAE S.L. BITING TIME

The peak biting period of the *Anopheles gambiae* s.l. was between 4:00 and 5:00 early in the morning inside in Lunga in Milenge after IRS. *Anopheles gambiae* s.l. started biting inside early in the evening between 7:00–8:00 pm and the peak was between 1:00 and 2:00 am in Shibesa in Mwense District. However, the biting rates are generally low and difficult to establish the trends for this species from the few collections of the human landing catches in most of the areas.

Figures 23, 24, 25, and 26 show the hourly biting rate of *Anopheles gambiae* s.l. per sprayed site after IRS.¹

Figure 23: *Anopheles gambiae* s.l. feeding time in Lunga (sprayed site) in Milenge

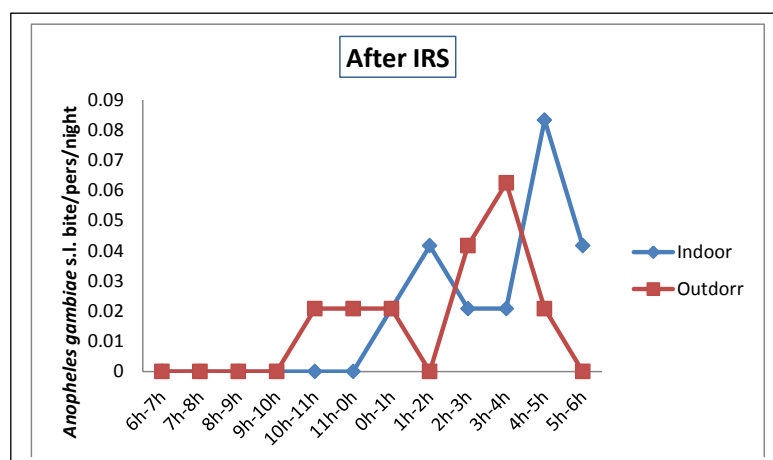


Figure 24: *Anopheles gambiae* s.l. feeding time in Shibesa (sprayed site) in Mwense

¹ *Anopheles gambiae* s.l. was not collected in Lunga in Milenge, in Kalonga, in Kasama and in Mbalani in Katete during the pre-spray period in August. Mosquito collection was also not performed in Shibesa in Mwense during the pre-spray period in August.

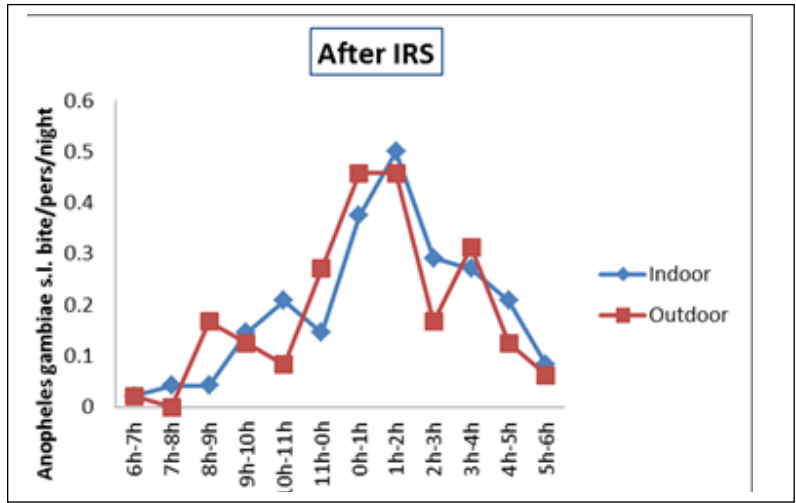


Figure 25: *Anopheles gambiae* s.l. feeding time in Kalonga (sprayed site) in Kasama

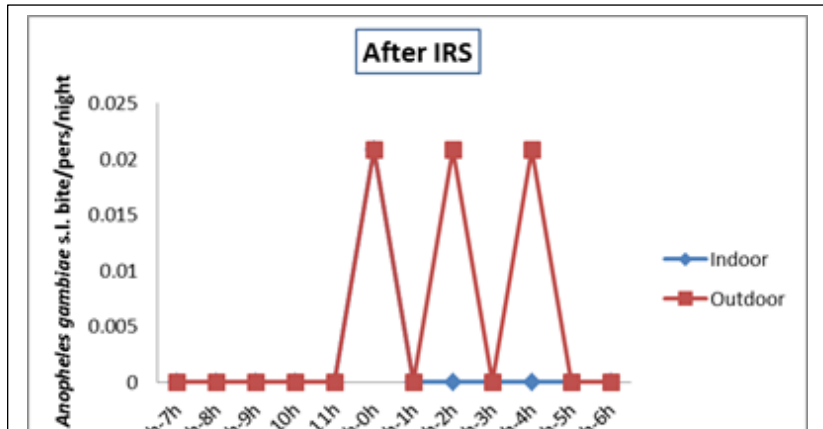
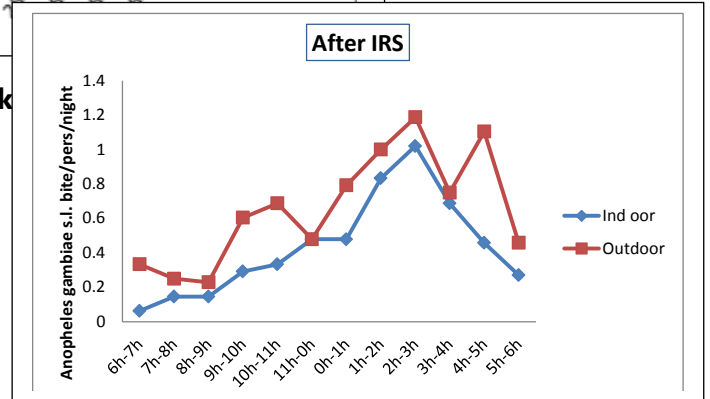
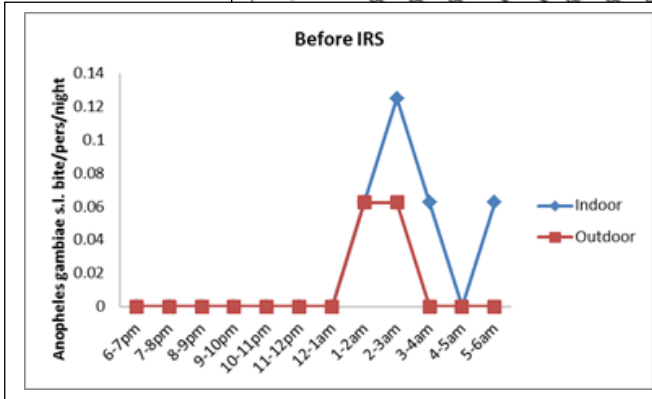


Figure 26:

Anopheles



3.5 PARITY RATE

The parity rate is similar both in sprayed and control sites. The overall parity rate for the IRS sites after IRS campaign was 45.5% (45/99) and 55.4% (243/439) for the control sites for *Anopheles funestus* s.l. and 54.3% (38/70) for IRS sites and 49.2% (31/63) for the control sites for *Anopheles gambiae* s.l. There is no statistically significant difference between the parity rate in the sprayed and control sites ($p=0.07$ for *Anopheles funestus* s.l. and $p=0.55$ for *Anopheles gambiae* s.l.), probably due to the low number of mosquitoes dissected for most of the sites.

Table 16: *Anopheles funestus* s.l. parity rate in Milenge and Mwense districts

Time	Lunga			Miyambo			Schibesa			Chebele		
	# <i>An. funestus</i> s.l. dissected	Parous	% parous	# <i>An. funestus</i> s.l. dissected	Parous	% parous	# <i>An. funestus</i> s.l. dissected	Parous	% parous	# <i>An. funestus</i> s.l. dissected	Parous	% parous
Pre-spray	4	2	50	51	41	80						
November	5	3	60	28	14	50	2	0	0	11	7	64
December	62	30	48.4	113	72	64	4	2	50	73	28	38
January	2	2	100	196	109	56	3	1	33	0	0	0
Total	73	37	50.7	388	236	60.8	9	3	33	84	35	42

Table 17: *Anopheles funestus* s.l. parity rate in Serenje and Katete districts

Time	Chibobo			Chishi			Mbalani			Robert		
	# <i>An. funestus</i> s.l. dissected	Parous	% parous	# <i>An. funestus</i> s.l. dissected	Parous	% parous	# <i>An. funestus</i> s.l. dissected	Parous	% parous	# <i>An. funestus</i> s.l. dissected	Parous	% parous
Pre-spray	0	0	0	5	3	60	0	0	0	0	0	
November	0	0	0	0	0	0	0	0	0	0	0	0
December	3	2	66.7	18	13	72	0	0	0	0	0	0
January	0	0	0	0	0	0	0	0	0	0	0	0
Total	3	2	66.7	23	16	69.6	0	0	0	0	0	0

Table 18: *Anopheles funestus* s.l. parity rate in Kasama and Isoka districts

Time	Nsalamba			Kalonga		
	# <i>An.</i>	Parous	% parous	# <i>An.</i>	Parous	% parous

	<i>funestus</i> s.l. dissected			<i>funestus</i> s.l. dissected		
Pre-spray	3	1	33	0	0	0
November	6	1	17	0	0	0
December	3	2	66.7	0	0	0
January	3	1	0	6	1	17
Total	15	5	33.3	6	1	16.7

Table 19: *Anopheles gambiae* s.l. parity rate in Milenge and Mwense districts

Time	Lunga			Miyambo			Schibesa			Chebele		
	# <i>An. gambiae</i> s.l. dissected	Parous	% parous	# <i>An. gambiae</i> s.l. dissected	Parous	% parous	# <i>An. gambiae</i> s.l. dissected	Parous	% parous	# <i>An. gambiae</i> s.l. dissected	Parous	% parous
Pre-spray	0	0	0	0	0	0						
November	0	0	0	0	0	0	3	1	33	1	0	0
December	0	0	0.0	0	0	0	7	2	29	42	22	52
January	2	2	100	4	4	100	17	9	53	16	5	31
Total	2	2	100.0	4	4	100.0	27	12	44	59	27	46

Table 20: *Anopheles gambiae* s.l. parity rate in Serenje and Katete districts

Time	Chibobo			Chishi			Mbalani			Robert		
	# <i>An. gambiae</i> s.l. dissected	Parous	% parous	# <i>An. gambiae</i> s.l. dissected	Parous	% parous	# <i>An. gambiae</i> s.l. dissected	Parous	% parous	# <i>An. gambiae</i> s.l. dissected	Parous	% parous
Pre-spray	0	0	0	5	0	0	0	0	0	0	0	
November	0	0	0	0	0	0	0	0	0	0	0	0
December	0	0	0.0	0	0	0						0
January	0	0	0	0	0	0	0	0	0	0	0	0
Total	0	0	0.0	5	0	0.0	0	0	0	0	0	0

Table 21: *Anopheles gambiae* s.l. parity rate in Kasama and Isoka districts

Time	Nsalamba			Kalonga		
	# <i>An. gambiae</i> s.l. dissected	Parous	% parous	# <i>An. gambiae</i> s.l. dissected	Parous	% parous
Pre-spray	11	3	27	0	0	0

November	4	1	25	0	0	0
December	23	18	78.3	2	2	0
January	11	3	0	1	0	0
Total	49	25	51.0	3	2	66.7

3.6 QUALITY ASSURANCE OF IRS AND INSECTICIDE DECAY RATE

The initial cone bioassay to determine the quality of spraying was carried out in six districts. The wall bioassay was conducted 24 hours after spraying in 40 randomly selected houses. The Kisumu strain of *Anopheles gambiae* s.s. available in the insectary in October was not enough to cover all six sentinel sites for the T0 cone bioassay tests due to the ant invasion and destruction of some mosquitoes in the insectary in July-August.

Anopheles funestus s.l. was collected using backpack aspirators from Milenge and Mwense districts. Previous surveys showed that *An. funestus* is most abundant in Milenge and Mwense districts. The WHO susceptibility test was done against pirimiphos-methyl CS (insecticide used for spraying) and showed 100% mortality. This known susceptible *Anopheles funestus* s.l. strain was used for the cone bioassay in Milenge, Mwense, and Serenje districts. Susceptible Kisumu strain (*Anopheles gambiae* s.s.) reared in the insectary at the NMCC was used for the cone bioassay tests in the remaining three districts, namely Katete, Isoka, and Kasama. The two most commonly found surface types in the areas, cement and mud walls, were tested.

The cone bioassay tests were done according to the WHO test procedure. During the tests, the number of mosquitoes knocked down after 30 minutes and dead after 24 hours were recorded. All the mosquitoes exposed were dead after the 24h holding period. The 100% mortality rate was noted at all sites at T0 and T1, respectively, 24 hours and one month after spraying, showing that the spraying was of good quality and pirimiphos-methyl CS was still effective one month after spray. Subsequent testing was done in December, January, February, March and April respectively, two, three, four, five and six months after the IRS campaign to determine the decay rate of insecticide applied on the walls. The mortality rate was 100% for both mud and cement sprayed walls except in Kasama District where the mortality rate in December was 96% for the mud and 98% for the cement sprayed walls.

In December 2015, two months after the IRS campaign, the insecticide applied continued to be effective and the WHO minimum threshold of effectiveness was met for both mud and cement walls. Less than 80% of susceptible mosquitoes exposed to the sprayed cement in Milenge (63%) and to the sprayed mud in Serenje (71%) were killed in January three months after the spraying. Pirimiphos-methyl was effective on both mud and cement in Kasama, Isoka, Katete, and Mwense in February four months after spraying. The test mortality rate was less than the 80% WHO threshold on the mud sprayed walls in Milenge and on the cement in Serenje. The residual life of pirimiphos-methyl in these two districts is short and needs to be investigated. Pirimiphos-methyl is still effective on both mud and cement sprayed walls five months after spraying in Mwense and Isoka districts. However the mortality rate was below 80% for mud sprayed walls in Kasama and Katete five months after spraying.

Figures 27 and 28 show the mortality rate at T0 (24 hours after spraying), at T1 (one month after in November), at T2 (two months after spraying in December), at T3 (three months after in January) at T4 (four months after spraying in February 2016), at T5 (five months after spraying in March 2016) and at T6 (Six months after spraying in April 2016).

Figure 27: Mortality of Kisumu susceptible strain of *An. gambiae* s.s. after 30 mins exposure to pirimiphos-methyl CS and 24H holding period at T0, T1, T2, T3, T4, T5 and T6 in Kasama, Isoka, and Katete

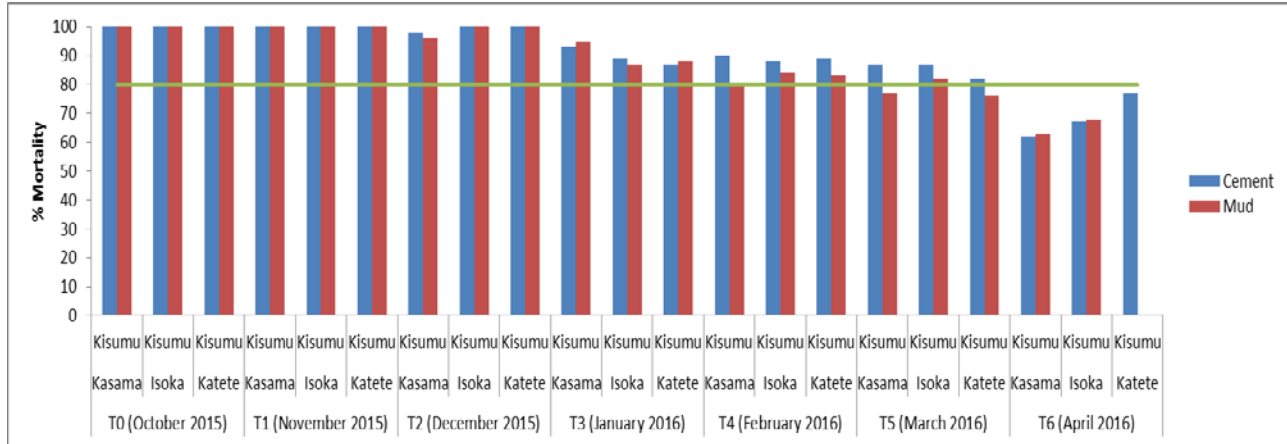
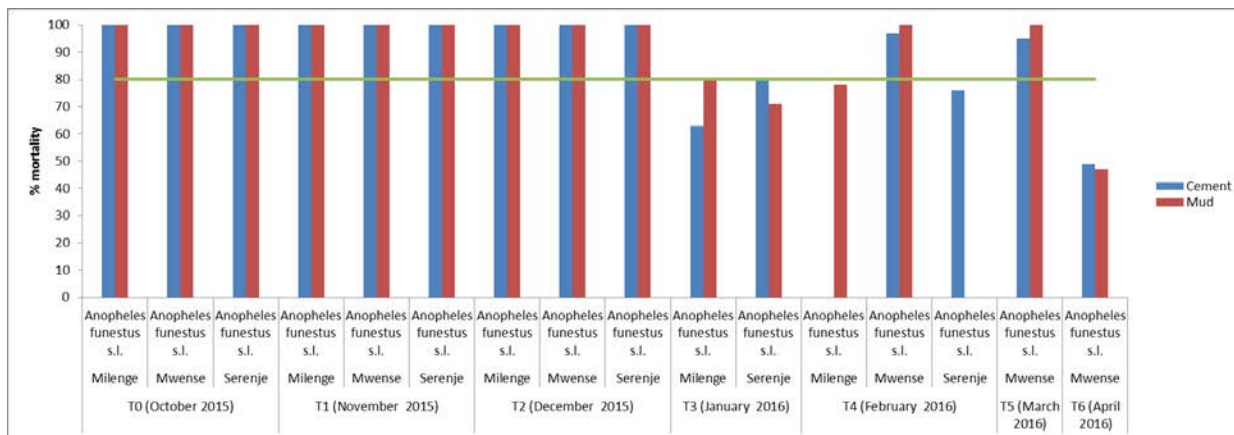


Figure 28: Mortality of *Anopheles funestus* s.l. susceptible strain after 30 mins exposure to pirimiphos-methyl CS and 24H holding period at T0, T1, T2, T3, T4, T5 and T6



3.7 MALARIA VECTOR SUSCEPTIBILITY TESTS

WHO tube test

An. funestus s.l. and *Anopheles gambiae* s.l. were fully susceptible to pirimiphos-methyl. A 100% mortality rate was recorded at all sites in 2015 and 2016. *Anopheles funestus* s.l. was resistant to deltamethrin at all sites but fully susceptible to Bendiocarb in Kalonga in Kasama and Chebele in Mwense districts. *Anopheles funestus* s.l. is resistant to Bendiocarb in Nandola in Kasama and in

Lunga and Miyambo in Milenge and in Chibobo in Serenje districts. *Anopheles funestus s.l.* is fully susceptible to DDT except in Mwemamusongo in Milenge.

Anopheles gambiae s.l. was fully susceptible to deltamethrin in Mbalani in Katete, in Chebele in Mwense, and in Lunga in Milenge and resistant in shibesa in Mwense and Robert in Katete.

Anopheles gambiae s.l. is fully susceptible to Bendiocarb at all sites but resistant to DDT in chebele in Mwense and in Lunga in Milenge.

Figure 29: Susceptibility status of *Anopheles funestus s.l.*

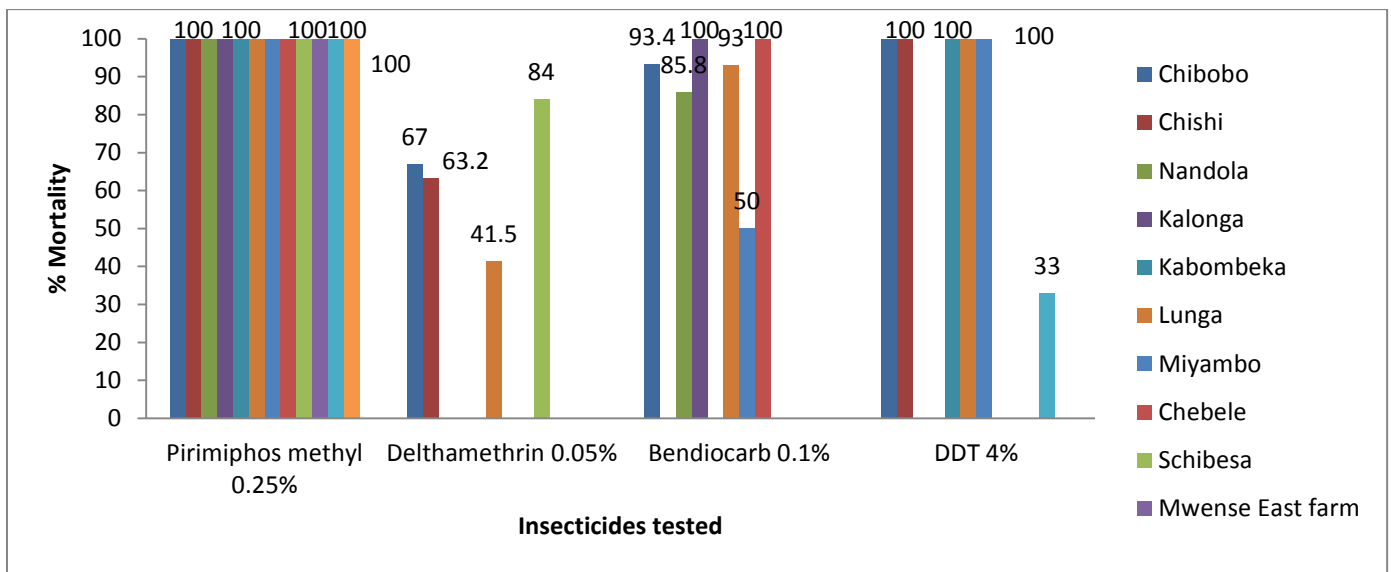
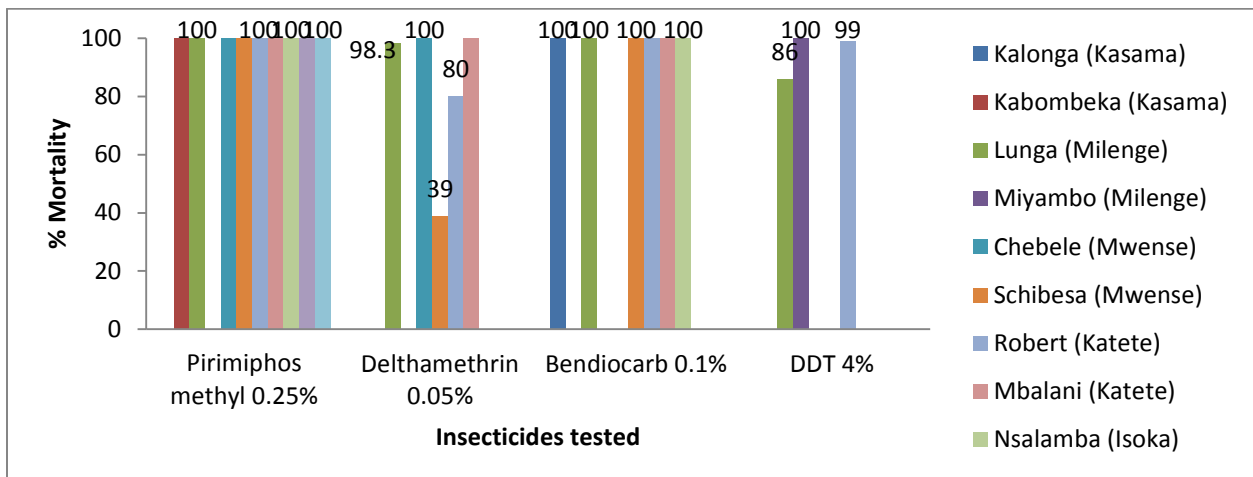


Figure 30 : Susceptibility status of *Anopheles gambiae s.l.*



Insecticide resistance intensity assays

The Rapid Resistance Intensity Diagnostic test (I-RDT) was performed in September and November with the adult female *Anopheles funestus* s.s. collected from seven villages in Milenge, two villages in Samfiya, and three villages in Mwense District. Additional tests were run from March to May 2016.

High intensity of resistance to pyrethroid was observed except for permethrin. *Anopheles funestus* s.s. collected from Fumpa Kalusa and Yatema was fully susceptible to five and ten times the diagnostic dose of permethrin. *Anopheles funestus* s.s. was resistant to deltamethrin at all selected sites in Milenge and Samfiya districts. No difference was observed between the sites in the intensity of deltamethrin resistance except in Shitambulli where 44% of the *Anopheles funestus* s.s. survived to ten times the diagnostic dose of deltamethrin. . 21% of *Anopheles funestus* s.s. from Miyambo exposed to five times the diagnostic dose of deltamethrin survived and 18% survived to 10 times the diagnostic dose. A difference was observed between the sites in the intensity of resistance for permethrin, (100%) of the *Anopheles funestus* s.s. exposed to five and ten times the diagnostic dose of Permethrin died in Fumpa Kalusa and Yatema. For Pwele village, 26% of the mosquitoes exposed survived to 10 times the diagnostic dose of deltamethrin. In addition, 14% of *Anopheles funestus* s.s. from Fumpa Kulusa survived to 10 times the diagnostic dose of deltamethrin.

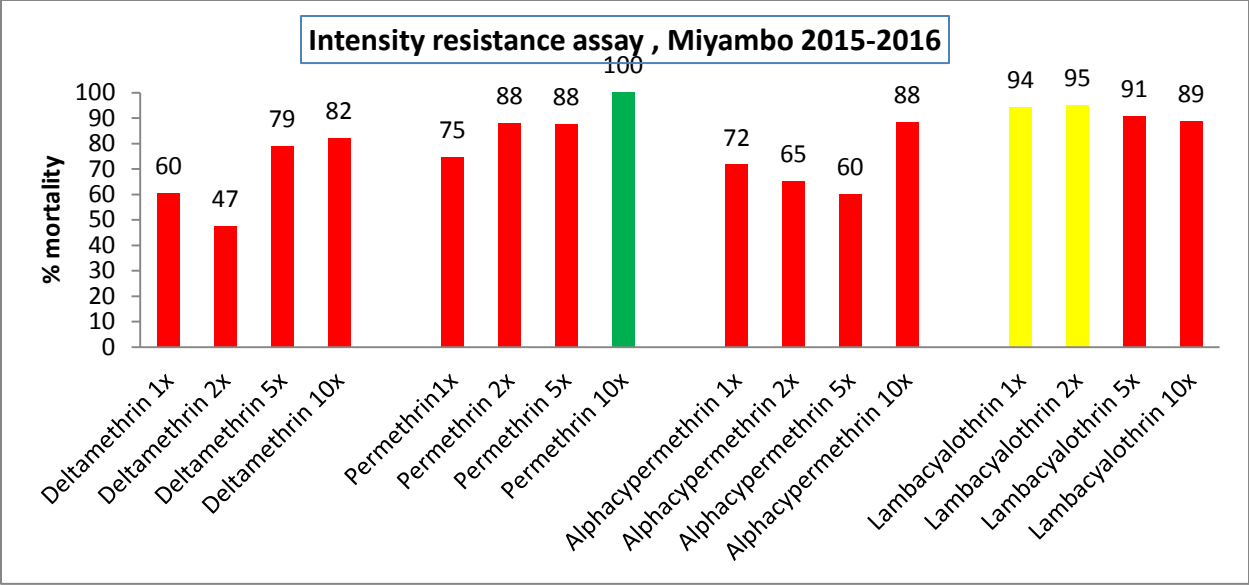
High intensity resistance is suspected in all the sites except in Shitambulli where the intensity of resistance seems moderate for deltamethrin.

Intensity resistance assay was performed in January for *Anopheles gambiae* s.l. but due to the heavy rain observed in January, few *Anopheles gambiae* were collected. Additional tests will be run during the next rainy season

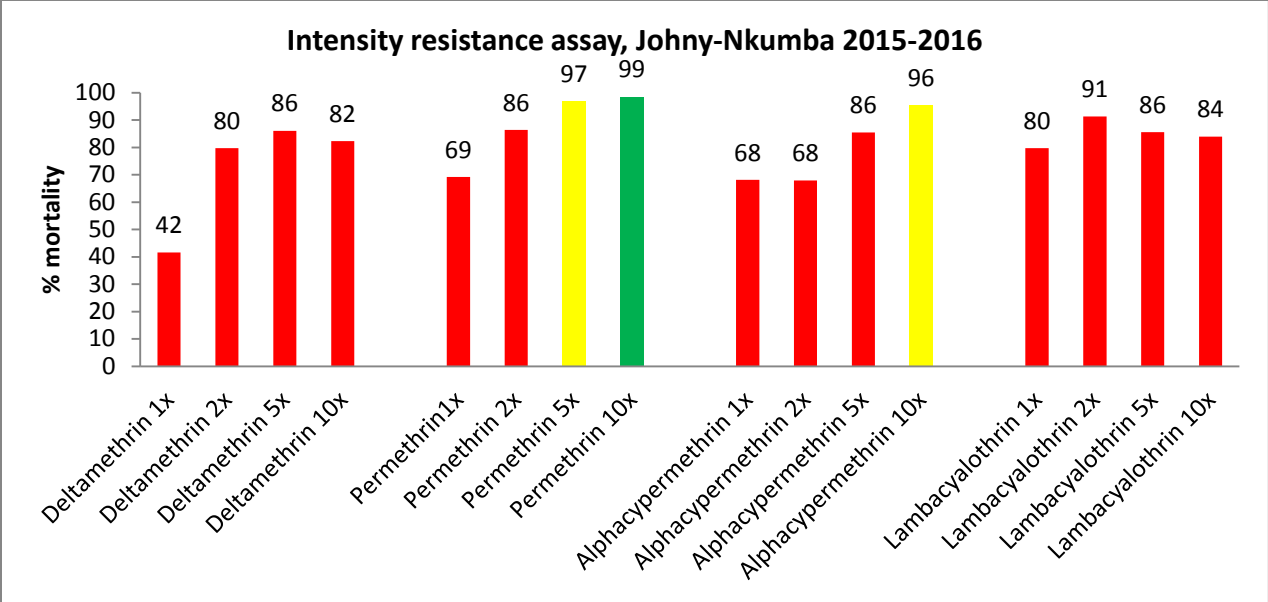
Figures 31 and 32 indicate the mortality rate per insecticide.

Figure 31: Intensity resistance assay in Milenge district against *An. funestus* s.l.

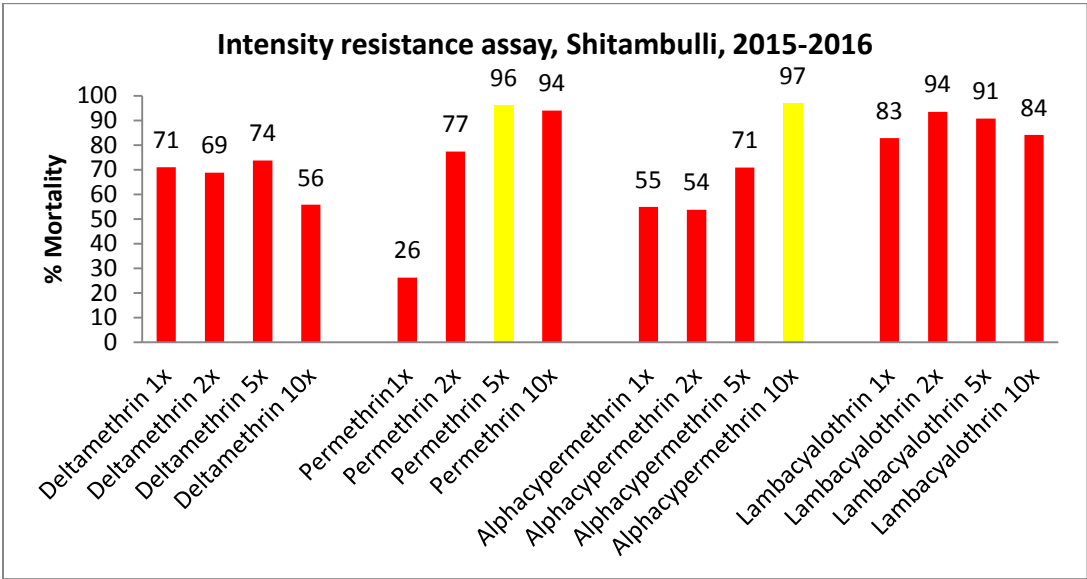
a)



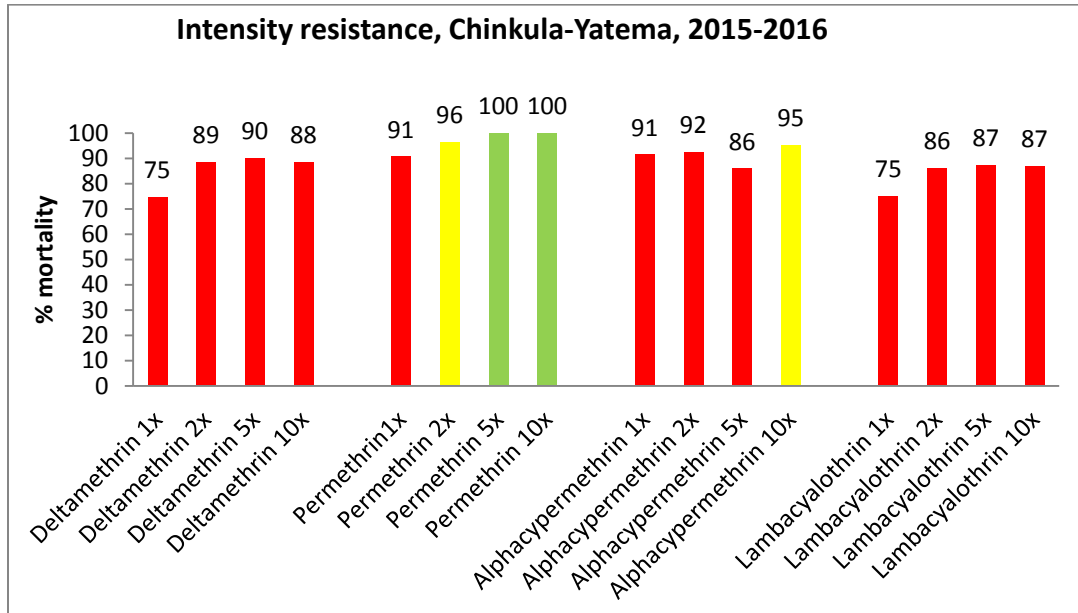
b)



c)



d)



e)

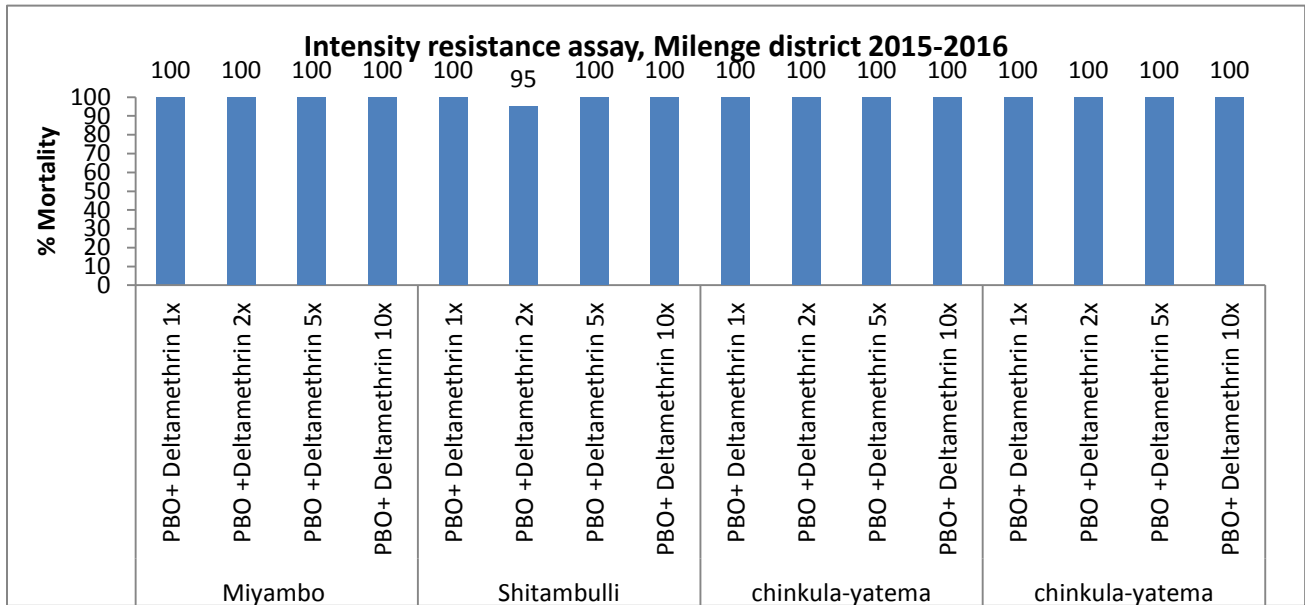
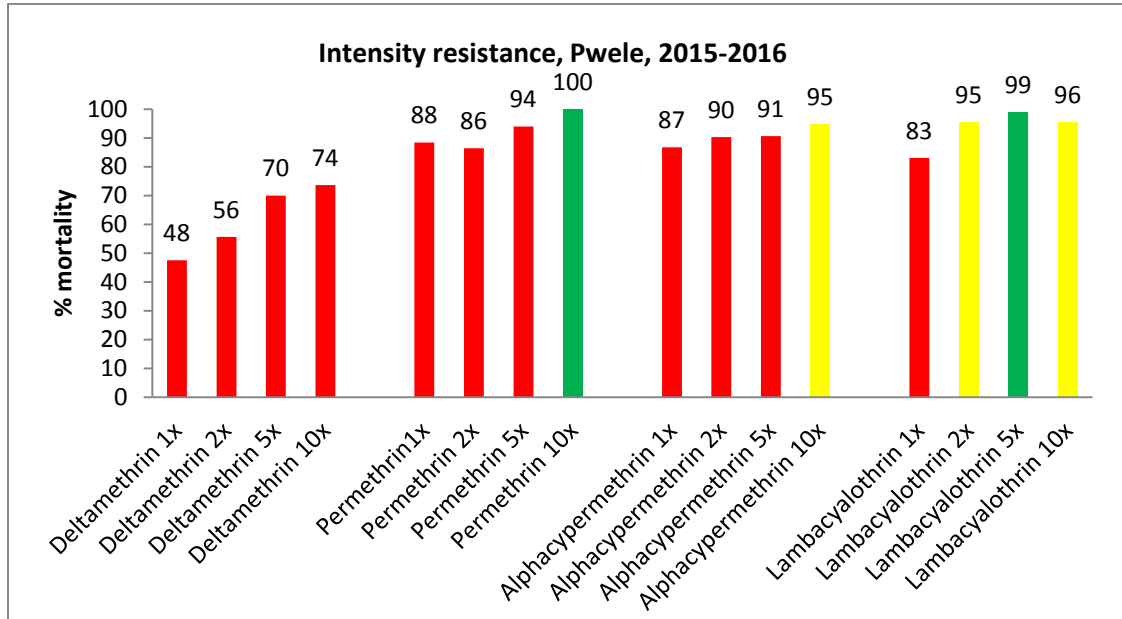
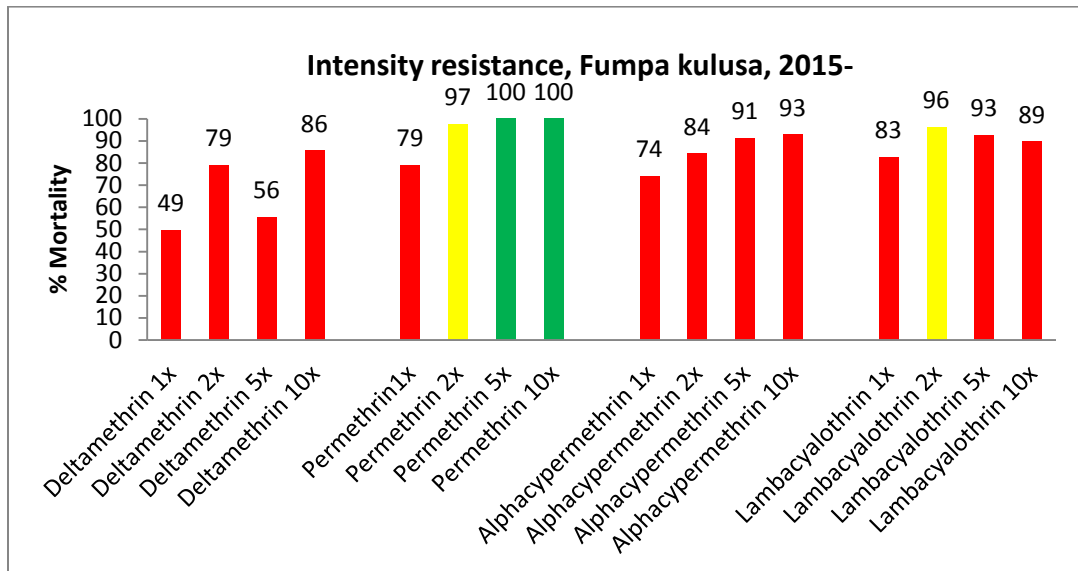


Figure 32: Intensity resistance assay in Samfya District

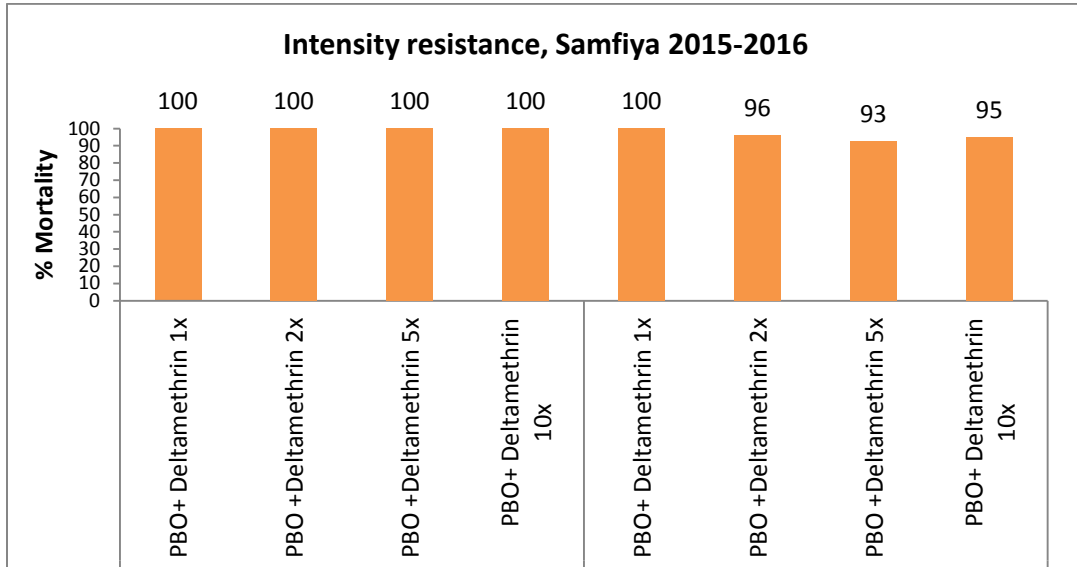
a)



b)



c)



4. TRAINING ON THE DISEASE DATA MANAGEMENT SYSTEM (DDMS)

The AIRS Zambia team, including the technical manager, the insectary technician, one field entomologist, the M&E manager, the database manager, and the IT specialist were trained from December 7 -11, 2015. AIRS Zambia started entering the data in the database but unfortunately is facing technical issues accessing to the DDMS using the office internet. The concern was shared with the DDMS focal person and the AIRS Zambia IT specialist.

5. COUNTRY CAPACITY BUILDING

AIRS Zambia has discussed and agreed with the National Malaria Control Program to establish new sentinel sites across the three epidemiological zones to create a representative sampling of geographical focus that will provide adequate entomological findings to inform planning for elimination interventions and strategies designed by the National Malaria Control Program and its partners.

Under ZISSP, 17 entomological kits were purchased, including microscope, backpack aspirators, larvae collections kits, and others entomology equipment. This equipment was sent to some districts but not used. AIRS Zambia assisted in retrieving these entomological surveillance field kits, which will be redistributed to the newly established sentinel sites, including the six PMI AIRS entomology sentinel districts.

Three Environmental Health Technicians were trained in 2015 by AIRS for entomology surveillance. Only one is involved in the entomology surveillance. Twelve technicians (two per sentinel site) were trained from March 2-4, 2016, on the mosquitoes sampling, the *Anopheles* species morphological identification using Gillies and M Coetzee, and 1987 identification key and ovary dissection. These trained staff will support the entomology surveillance at the district level.

6. CONCLUSION

The cone bioassay test conducted at T0, 24h and at T1, one month after spraying, showed 100% mortality of the mosquitoes exposed. The residual life of pirimiphos-methyl was less than six months at all sites. The vector density and the average HBR significantly decreased one and two months after spraying in the intervention sites compared to the control site and the baseline. An increase of the HBR was observed in January three months after the IRS campaign.

An. gambiae s.l. and *Anopheles funestus* s.l. are fully susceptible to pirimiphos-methyl. *Anopheles funestus* s.l. is resistant to deltamethrin at all sites; fully susceptible to Bendiocarb in Kalonga in Kasama and Chebele in Mwense. *Anopheles funestus* s.l. is resistant to Bendiocarb in Nandola in Kasama, in Lunga and Miyambo in Milenge, in Chibobo in Serenje. *Anopheles funestus* s.l. are fully susceptible to DDT except in Miyambo in Milenge. *Anopheles gambiae* s.l. was fully susceptible to deltamethrin in Mbalani in Katete, in Chebele in Mwense, and in Lunga in Milenge and resistant in Shibesa in Mwense and Robert in Katete. *Anopheles gambiae* s.l. is fully susceptible to Bendiocarb at all sites but resistant to DDT in chebele in Mwense and in Lunga in Milenge.

The resistance intensity assay shows that *Anopheles funestus* s.l. was resistant to all the four pyrethroids tested at all selected sites in Milenge and Samfiya districts. No difference was observed between the sites in the intensity of deltamethrin resistance in the area except in Shitambulli. However a difference was observed between the sites in the intensity of permethrin resistance.

The parity rate is similar in both sprayed and control sites. There is no statistically significant difference between the parity in the sprayed and control sites. The parity is high at most of the sites.

7. RECOMMENDATIONS

- *Anopheles gambiae* Kisumu strain should be used for the cone bioassay for the upcoming IRS campaign.
- In order to maximize our impact on entomological indicators, the focus for 2016 IRS season will be to continue to improve the quality of spraying by implementing DOS (directly observed spraying) by Team Leaders during the campaign.
- Adjust timing of spraying to start in November in the Eastern Province so it's in line with the peak of *Anopheles funestus* s.l. and the decay of pirimiphos methyl.

ANNEX

Table A: Culicidae collected by PSC and CDC light trap in August before IRS campaign

Districts	Villages	Status	CDC light trap						Pyrethrum Spray Catch		
			<i>An.funestus</i> <i>s.l</i>	<i>An.gambiae</i> <i>s.l</i>	<i>An.</i> <i>tenebrosus</i>	<i>An.</i> <i>tchedi</i>	<i>An.</i> <i>squamosus</i>	<i>Culicinae</i>	<i>An.</i> <i>funestus</i> <i>s.l</i>	<i>An</i> <i>.gambiae</i> <i>s.l</i>	<i>Culicinae</i>
			n	n	n	n	n	n	n	n	n
Kasama	Kalunga	Sprayed	1	0	0	0	0	0	0	0	3
Katete	Mbalani	Sprayed	0	0	0	0	0	13	0	0	27
	Robert	Control	0	0	5	0	0	0	0	0	0
Isoka	Nsalamba	Sprayed	1	2		1	0	16	4	0	10
Mwense	Shibesa	Sprayed	233	3	0	0	0	7	58	0	4
	Chebele	Control	66	2	0	0	0	11	17	1	6
Milenge	Lunga	Sprayed	213	0	161	46	0	1	224	0	0
	Niyambo	Control	378	0	18	8	1	6	74	0	0
Serenje	chibobo	Sprayed	1	0	0	0	0	0	0	0	0
	Chichi	Control	4	0	0	0	0	0	1	0	0
Total			897	7	184	55	1	54	378	1	50

Table B: Culicidae collected by Human Landing Catch in August 2015 before IRS campaign

Districts	Villages	Status	HLC											
			Indoor						Outdoor					
			<i>An. funestu</i> s.l.	<i>An.gambia</i> e s.l.	<i>An. tenebrosu</i> s	<i>An. tcheke</i> di	<i>An. squamosu</i> s	<i>culicin</i> e	<i>An. funestu</i> s.l.	<i>An.gambia</i> e s.l.	<i>An. tenebrosu</i> s	<i>An. tcheke</i> di	<i>An. squamosu</i> s	<i>Culicin</i> e
			n	n	N	n	n	n	n	n	n	n	n	n
Kasama	Kalonga	Sprayed	6	0	7	0	0	133	1	0	0	1	0	55
Katete	Mbalani	Sprayed	0	0	0	0	0	206	0	0	0	0	0	166
	Robert	Control	1	0	12	0	0	2	0	0	1	0	0	0
Isoka	Nsalamba	Sprayed	2	5	3	2	5	3	11	2	3	6	19	6
Mwense	Shibesa	Sprayed												
	Chebele	Control												
Milenge	Lunga	Sprayed	220	0	330	20	6	8	93	0	760	28	23	34
	Niyambo	Control	497	0	170	12	5	3	202	0	372	29	7	5
Serenje	chibobo	Sprayed	0	0	0	0	0	0	0	0	0	0	0	0
	Chichi	Control	5	0	0	0	0	0	3	0	0	0	1	0
Total			731	5	522	34	16	355	310	2	1136	64	50	266

Table C: Culicidae collected by PSC and CDC light trap in November 2015

Districts	Villages	Status	CDC light trap						Pyrethrum Spray Catch		
			<i>An.funestus s.l.</i>	<i>An.gambiae s.l.</i>	<i>An. tenebrosus</i>	<i>An. tchekeledi</i>	<i>An. squamosus</i>	<i>Culicinae</i>	<i>An. funestus s.l.</i>	<i>An .gambiae s.l.</i>	<i>Culicinae</i>
			n	N	n	n	n	n	n	n	n
Kasama	Kalonga	Sprayed	0	0	0	0	0	1	0	0	15
Katete	Mbalani	Sprayed	0	0	0	0	0	15	0	0	8
	Robert	Control	0	0	0	0	0	24	0	0	83
Isoka	Nsalamba	Sprayed	1	11	0	6	4	12	2	1	18
Mwense	Shibesa	Sprayed	16	1	0	0	0	3	1	0	0
	Chebele	Control	610	6	0	0	0	0	89	0	0
Milenge	Lunga	Sprayed	49	0	1	1	0	12	393	0	9
	Niyambo	Control	165	0	0	0	0	2	568	0	3
Serenje	chibobo	Sprayed	0	0	0	0	0	0	40	0	0
	Chichi	Control	3	0	0	0	0	0	28	0	0
Total			844	18	1	7	4	69	1121	1	136

Table D: Culicidae collected by Human Landing Catch in November 2015

Distri cts	Villag es	Statu s	HLC															
			Indoor								Outdoor							
			<i>An. funes tus s.l.</i>	<i>An.gam biae s.l.</i>	<i>An. tenebr osus</i>	<i>An. tchek edii</i>	<i>An. squam osus</i>	<i>An. Coust ani</i>	<i>An. rufi pes</i>	<i>Culic ine</i>	<i>An. funes tus s.l.</i>	<i>An.gam biae s.l.</i>	<i>An. tenebr osus</i>	<i>An. tchek edii</i>	<i>An. squam osus</i>	<i>An. coust ani</i>	<i>An. rufi pes</i>	<i>Culic ine</i>
			n	n	n	N	n	n	n	n		n	n	n	n	n	n	n
Kasa ma	Kalong a	Spray ed	0	0	0	1	1	0	0	8	1	0	9	1	2	0	0	30
Katet e	Mbala ni	Spray ed	0	0	0	0	0	0	0	5	0	0	0	0	0	0	0	6
	Robert	Contr ol	0	0	0	0	0	0	0	27	0	0	0	0	0	0	0	11
Isoka	Nsala mba	Spray ed	1	15	0	8	1	0	0	13	13	18	0	20	28	0	1	160
Mwe nse	Shibes a	Spray ed	4	5	0	0	0	0	0	3	1	2	0	0	0	0	1	22
	Chebel e	Contr ol	960	70	2	0	0	1	0	4	510	100	9	0	1	22	0	29
Milen ge	Lunga	Spray ed	36	0	96	42	0	0	0	31	17	0	102	67	0	0	0	69
	Niyam bo	Contr ol	260	0	29	7	0	0	0	21	91	0	86	8	0	0	0	82
Seren je	chibob o	Spray ed	3	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0
	Chichi	Contr ol	1	0	0	0	0	0	0	5	1	0	0	0	0	0	0	3
Tota l			1265	90	127	58	2	1	1	117	634	120	206	96	31	22	2	412

Table E: Culicidae collected by PSC and CDC light trap in December 2015

Districts	Villages	Status	CDC light trap						Pyrethrum Spray Catch		
			<i>An.funestus</i> s.l.	<i>An.gambiae</i> s.l.	<i>An.</i> <i>tenebrosus</i>	<i>An.</i> <i>tchekei</i>	<i>An.</i> <i>squamosus</i>	<i>Culicinae</i>	<i>An.</i> <i>funestus</i> s.l.	<i>An</i> <i>.gambiae</i> s.l.	<i>Culicinae</i>
			n	N	n			N	n	n	n
Kasama	Kalonga	Sprayed	2	0	0	0	0	12	6	1	44
Katete	Mbalani	Sprayed	0	0	0	0	0	25	0	0	22
	Robert	Control	0	0	0	0	0	4	0	0	50
Isoka	Nsalamba	Sprayed	7	25	0	2	4	46	1	3	0
Mwense	Shibesa	Sprayed	39	6	0	0	0	3	2	4	2
	Chebele	Control	40	6	0	0	0	16	64	8	2
Milenge	Lunga	Sprayed	65	0	8	0	0	9	65	0	0
	Niyambo	Control	296	0	0	0	0	3	243	0	0
Serenje	chibobo	Sprayed	5	0	0	0	0	0	27	0	3
	Chichi	Control	36	0	0	0	0	3	93	0	3

Total			490	37	8	2	4	121	501	16	126
--------------	--	--	------------	-----------	----------	----------	----------	------------	------------	-----------	------------

Table F: Culicidae collected by Human Landing Catch in December 2015

Districts	Villages	Status	HLC																	
			Indoor									Outdoor								
			An. funestus s.l.	An.gambiae s.l.	An. tenebrosus	An. tchekeidii	An. squamosus	An. Coustani	An. rufipes	An. ziemanni	Culicine	An. funestus s.l.	An.gambiae s.l.	An. tenebrosus	An. tchekeidii	An. squamosus	An. coustani	An. rufipes	An. Ziemanni	Culicine
Kasama	Kalongo	Sprayed	8	1	42	3	11	0	0	0	13	2	3	68	6	8	0	0	0	19
Katete	Mbalani	Sprayed	0	0	0	0	0	0	0	0	284	0	0	0	0	0	0	0	0	118
	Robert	Control	1	1	0	0	0	0	0	0	22	1	0	0	0	0	0	0	0	13
Isoka	Nsalamba	Sprayed	2	8	1	4	2	0	0	0	28	7	44	2	25	10	0	0	0	247
Mwenese	Shibesa	Sprayed	18	27	0	0	1	0	0	0	6	31	36	0	0	1	0	0	0	348
	Chebele	Control	179	52	0	0	0	0	0	0	14	100	68	1	0	2	0	0	0	45
Mileenge	Lunga	Sprayed	126	0	24	20	0	0	0	7	47	79	1	41	25	0	0	0	17	77
	Niyambo	Control	759	2	11	3	0	1	0	1	39	397	0	39	8	1	0	0	2	176

Sere nje	chibo bo	Spra yed	6	0	0	0	0	0	0	0	0	15	0	0	0	1	1	0	0	10
	Chichi	Cont rol	82	0	0	0	0	2	0	0	3	47	0	0	0	1	6	0	0	22
Tota l			118 1	91	78	30	14	3	0	8	456	679	152	151	64	24	7	0	19	107 5

Table G: Culicidae collected by PSC and CDC light trap in January 2016

District s	Villages	Status	CDC light trap							Pyrethrum Spray Catch						
			<i>An.funest us s.l.</i>	<i>An.gambi ae s.l.</i>	<i>An. tenebros us</i>	<i>An. tcheke dii</i>	<i>An. squamos us</i>	<i>An. cousta ni</i>	<i>Culicin ae</i>	<i>An. funest us s.l.</i>	<i>An .gambi ae s.l.</i>	<i>An. squamos us</i>	<i>An. tcheke dii</i>	<i>An. cousta ni</i>	<i>An. tenebros us</i>	<i>Culicin ae</i>
			n	n	n	n	n		n	n	n		n			n
Kasam a	Kalonga	Spra yed	6	6	1	3	12	0	17	9	0	n	0	0	0	49
Katete	Mbalani	Spra yed	0	0	0	0	0	0	223	0	0	0	0	0	0	76
	Robert	Contro l	1	0	0	0	0	0	19	0	1	0	0	0	0	8
Isoka	Nsalam ba	Spra yed	9	231	1	4	13	0	33	14	18	2	1	0	0	12
Mwens e	Shibesa	Spra yed	60	73	0	0	0	2	76	41	51	0	0	0	0	4

	Chebele	Control	15	43	0	0	0	0	0	115	11	39	0	0	2	0	3
Milenge	Lunga	Sprayed	206	24	4	0	0	0	0	33	119	1	0	0	0	0	0
	Niyambo	Control	735	12	182	4	2	0	0	143	155	0	2	0	0	19	33
Serenje	chibobo	Sprayed	1	0	0	0	0	0	0	26	34	0	0	0	0	0	1
	Chichi	Control	38	0	0	0	0	0	0	17	108	0	0	0	0	0	0
Total			1071	389	188	11	27	2	702	491	110	4	1	2	19	186	

Table H: Culicidae collected by Human Landing Catch in January 2016

Districts	Villages	Status	HLC																	
			Indoor									Outdoor								
			<i>An. funestus</i> s.l.	<i>An.gambiae</i> s.l.	<i>An. tenebrosus</i>	<i>An. tcheke</i> edii	<i>An. squamosus</i>	<i>An. Coustani</i>	<i>An. rufipes</i>	<i>An. ziemani</i>	<i>Culicine</i>	<i>An. funestus</i> s.l.	<i>An.gambiae</i> s.l.	<i>An. tenebrosus</i>	<i>An. tcheke</i> edii	<i>An. squamosus</i>	<i>An. coustani</i>	<i>An. rufipes</i>	<i>An. ziemani</i>	<i>Culicine</i>
Kasama	Kalonga	Sprayed	31	6	31	39	75	80	0	0	6	16	3	67	90	211	236	0	0	18
Katete	Mbalani	Sprayed	0	0	0	0	0	0	0	0	460	0	0	0	0	0	0	0	0	130
	Robert	Control	1	7	1	0	0	0	0	0	44	0	0	0	0	0	0	0	0	12

		ol																		
Isoka	Nsalamba	Sprayed	22	222	15	31	47	3	0	0	106	40	332	42	83	219	31	0	0	470
Mwenese	Shibesa	Sprayed	49	80	0	0	0	0	0	0	66	40	70	0	0	0	14	0	0	31
	Chebele	Control	44	151	0	0	0	9	0	0	95	36	164	0	0	0	21	0	0	30
Milenge	Lunga	Sprayed	327	11	131	12	0	0	0	0	217	176	8	185	14	2	0	0	0	317
	Niyambo	Control	828	7	50	1	0	0	0	0	159	483	11	95	0	1	0	0	0	196
Serenje	chibobo	Sprayed	12	0	0	0	0	1	0	0	3	10	0	0	0	0	2	0	0	0
	Chichi	Control	40	0	0	0	0	0	0	0	9	10	0	0	0	0	3	0	0	5
Total			1354	484	228	83	122	13	0	0	1165	811	588	389	187	433	307	0	0	1209

Table I: Number of *Anopheles gambiae* s.s. Kisumu strain exposed and dead in Kasama, Isoka and Katete

District	Type of wall	October			November			December			January			February			Observations
		Number exposed	Number of dead at the end of the 24 hours	% of mortality observed at the end of the 24H	Number exposed	Number of dead at the end of the 24 hours	% of mortality observed at the end of the 24H	Number exposed	Number of dead at the end of the 24 hours	% of mortality observed at the end of the 24H	Number exposed	Number of dead at the end of the 24 hours	% of mortality observed at the end of the 24H	Number exposed	Number of dead at the end of the 24 hours	% of mortality observed at the end of the 24H of holding period	

			holding period	of holding period		holding period	of holding period		holding period	of holding period		holding period	of holding period		holding period	
Kasama	Mud	60	60	100	60	60	100	60	58	97	61	58	95	60	48	80
	Cement	60	60	100	60	60	100	60	59	98	61	57	93	60	54	90
Isoka	Mud	60	60	100	60	60	100	60	60	100	60	52	87	60	51	84
	Cement	60	60	100	61	61	100	60	60	100	63	56	89	60	53	87
Katete	Mud	61	61	100	60	60	100	60	60	100	60	53	88	63	52	82.53968
	Cement	60	60	100	60	60	100	60	60	100	60	52	87	62	55	88.70968

Mortality in the control was 7.5%. Abbott formula was used

Table J: Number of *Anopheles funestus* s.l. susceptible strain exposed and dead per site and per month in Milenge, Mwense and Serenje

District	Type of wall	October			November			December			January			February		
		Number exposed	Number of dead at the end of the 24 hours holding period	% of mortality observed at the end of the 24H of holding period	Number exposed	Number of dead at the end of the 24 hours holding period	% of mortality observed at the end of the 24H of holding period	Number exposed	Number of dead at the end of the 24 hours holding period	% of mortality observed at the end of the 24H of holding period	Number exposed	Number of dead at the end of the 24 hours holding period	% of mortality observed at the end of the 24H of holding period	Number exposed	Number of dead at the end of the 24 hours holding period	% of mortality observed at the end of the 24H of holding period
Milenge	Mud	92	92	100	90	90	100	90	90	100	90	72	80	60	47	78

	Cement	91	91	100	90	90	100	91	91	100	90	57	63			
Mwense	Mud	90	90	100	95	95	100	90	90	100				60	60	100
	Cement	90	90	100	93	93	100	90	90	100				60	59	98
Serenje	Mud	95	95	100	90	90	100	94	94	100	90	64	71			
	Cement	93	93	100	90	90	100	97	97	100	90	72	80	62	47	76

Table K: Susceptibility status of *An. funestus* s.l.

District	Pirimiphos-Methyl 0.25%				Deltamethrin 0.05%		Bendiocarb 0.1%		DDT 4%			
	February-August 2015		October 2015-March 2016		February-August 2015		February-August 2015		February-August 2015		October 2015-March 2016	
	n	%M	n	%M	n	%M	n	%M	n	%M	n	%M
Kasama (Nandola)	100	100					99	85.8				
Kasama (Kalonga)	104	100					50	100				
Kabombeka (Kasama)			24	100					5	100		
Milenge (Lunga)	138	100	20	100	41	41.5	100	93	50	100		
Milenge (Miyambo)	15	100					20	50			8	100
Serenje (chibobo)	65	100			64	67	60	90	144	100		
Serenje (Chishi)	112	100			101	63.2			17	100		
Mwense (Chebele)	33	100					31	100				

Mwense (Shibesa)	31	100			74	84						
Mwense (East farm)			50	100								
Milenge (Mwemamusongo)			19	100								
Katete (Robert)			40	100								

Table L: Susceptibility status of *An. gambiae* s.l.

District	Pirimiphos-Methyl 0.25%				Deltamethrin 0.05%		Bendiocarb 0.1%		DDT 4%					
	February-August 2015		October 2015-March 2016		April- June 2016		March- June 2016		March- June 2016		October 2015-March 2016		April- June 2016	
	n	%M	n	%M	n	% M	n	% M	n	% M	n	% M	n	% M
Kasama (Kalonga)									25	100	23	0		
Kabombeka (Kasama)			39	100										
Milenge (Lunga)					94	100	60	98.3	78	100			50	86
Milenge (Miyambo)													23	100
Mwense (chebele)	75	100					6	100			102	41		
Mwense (Shibesa)			64	100			72	39	50	100				
Katete (Robert)					146	100	122	80	136	100			136	99

Katete (Mbalani)					66	100	26	100	63	100				
Isoka (Nsalamba)					73	100			29	100				
Isoka (Chilanga)					23	100								
Isoka (Londamaka)					18	100								